

This Page Is Inserted by IFW Operations
and is not a part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representation of
The original documents submitted by the applicant.

Defects in the images may include (but are not limited to):

- BLACK BORDERS
- TEXT CUT OFF AT TOP, BOTTOM OR SIDES
- FADED TEXT
- ILLEGIBLE TEXT
- SKEWED/SLANTED IMAGES
- COLORED PHOTOS
- BLACK OR VERY BLACK AND WHITE DARK PHOTOS
- GRAY SCALE DOCUMENTS

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images,
please do not report the images to the
Image Problem Mailbox.**

THIS PAGE BLANK (USPTO)

(9)



Europäisches Patentamt
European Patent Office
Office européen des brevets

(11) Publication number:

**0 319 957
A2**

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 88120466.3

(51) Int. Cl.⁴: **C08J 7/12 , C12N 11/00 ,
G01N 33/543 , C12Q 1/68**

(22) Date of filing: 07.12.88

(30) Priority: 07.12.87 DK 6414/87

(43) Date of publication of application:
14.06.89 Bulletin 89/24

(84) Designated Contracting States:
ES GR

(71) Applicant: **GLUETECH APS**
Copenhagen Science Park Symbion
Haraldsgade 68
DK-2100 Copenhagen Ø(DK)

(72) Inventor: **Elsner, Henrik**
Svend Gongsvej 36
DK-2700 Bronshøj(DK)
Inventor: **Mouritsen, Søren**
Vandtaarnsvej 7, lejl. 81
DK-3460 Birkerød(DK)

(74) Representative: **Patentanwälte TER MEER -**
MÜLLER - STEINMEISTER
Mauerkircherstrasse 45
D-8000 München 80(DE)

(54) **A method for modifying the surface of a polymer.**

(57) The surface of a polymer may be modified by contacting the polymer with a two or three membered heterocyclic compound I under electromagnetic irradiation. The heterocyclic compound may be substituted and may comprise linking units and/or probes so as to render the combination of polymer and compound I able to bind biomolecules and/or to be identified. Compounds I are preferably substituted psoralens, and the modified polymers can be used in e.g. ELISA methods, bioreactors, chromatography, solid phase chemical synthesis or bio-sensors.

EP 0 319 957 A2

A METHOD FOR MODIFYING THE SURFACE OF A POLYMER

BRIEF DESCRIPTION OF THE INVENTION

The present invention relates to a method for modifying the surface of a polymer. In particular, the invention relates to a method for immobilizing compounds on a solid polymer phase, preferably polystyrene, polyvinyl chloride or polyethylene terephthalate glycol.

BACKGROUND OF THE INVENTION

Immobilization of biomolecules on solid phases are widely used in numerous techniques as e.g. chromatography, in bio-sensors, in bio-reactors for e.g. solid phase enzyme processing, chemical synthesis of peptides, oligonucleotides or other compounds and in so-called heterogeneous immunoassays.

In the heterogeneous immunoassays, antigens or antibodies can be covalently coupled to carriers such as cellulose, agarose or polyacrylamide. However, when the compounds are to be bound on a solid phase, usually polystyrene, polypropylene or polyvinylchloride test tubes or micro-titre-plates, physical adsorption of the compounds has been the normal coupling method in heterogeneous immunoassays (Engvall and Pearlmann, J. Immunol., 1972, p. 109-129).

Passive physical adsorption is, however, not irreversible (Lehtonen et al., J. Immunol. Methods, 1980, p. 34-61), which may affect the reproducibility of the immunoassays, especially when the antigen/antibody coated solid phase is stored in dry form. Also solid phase adsorbed antigen may not be recognized by its corresponding antibody because of denaturation (reconformation) of the antigen tertiary structure (Kurki and Virtanen, J. Immunol. Methods, 1984, p. 67-122). The antigen may also exhibit new antigenic determinants when denatured, as in the case of DNA.

The ability of passive binding to plastic is furthermore restricted to a limited amount of molecules as e.g. proteins, or single-stranded DNA. However, some proteins and nucleic acids as well as polysaccharides and smaller molecules can not be adsorbed directly to some types of plastic.

Covalent binding, in contrast to simple physical binding, orientates all immobilized compounds in a defined way on the solid phase, thereby exposing defined areas on e.g. antigens, antibodies or enzyme catalytic sites to the fluid phase eventually poured on the solid phase surface. Antigen epitopes or active sites on these compounds can in this way remain functional.

Irreversible immobilization of molecules may furthermore have some advantages in relation to storage of solid phase immobilized compounds in their dry state, since passive physical binding partly denatures e.g. some proteins and nucleic acids.

For all these reasons it would be advantageous if it was possible to immobilize antigens, antibodies or other molecules covalently to the solid phase.

Covalent solid phase fixation of some types of compounds to polystyrene has been known for several years. Solid phase peptide and oligonucleotide synthesis are e.g. performed using modified polystyrene particles as the solid support. However, these modification methods frequently involve very hazardous chemicals and several time-consuming operation steps. An example of this is the preparation of Merrifield's Peptide Resin which has been widely used in peptide synthesis. Preparation of this resin involves the extremely carcinogenic reagent chloromethylmethyl ether and this and other organic solvents often result in a turbid surface because the plastic is slightly soluble in the reagents. This is inconvenient if a subsequent spectrophotometrical detection is desired.

Polystyrenes premodified with e.g. -OH, -SO₃H or -NH₂ groups are available. When using these premodified polystyrenes a separate production of particles for each type of modified polystyrene is necessary.

A method for introducing amino groups on plastic surfaces for the use in micro-titre-plates has been described (J. Virol. Methods 3, 1981, p. 155-165). This procedure, which requires the hazardous chemicals methanesulphonic acid, glacial acetic acid and fuming nitric acid, takes two days and needs a well functioning hood. Such conditions can hardly be used in large scale production due to environmental demands. Furthermore, these methods are generally time-consuming and it is difficult to obtain a uniform surface modification.

Chemical modification of polymer materials usually results in a heterogeneous mixture of products on the polymer surface since it is not possible to separate the solid phase bound main functional groups from the

unwanted products of the reaction. This results more or less in a mixture of different active groups with different binding specificities to e.g. proteins.

Another method of chemically modifying or activating polymer surfaces by introducing functional groups such as OH, NH₂, COOH, CHO, NCO or SCO is suggested in EP A patent application no. 155.252.

5 According to this application a solid polymer surface is activated by radiation grafting of vinyl monomers having at least one functional group capable of binding to biologically active molecules. The polymer surface is grafted in solution in a chain transfer reducing solvent at a very low monomer concentration not exceeding 3% by weight when the vinyl monomer is 1-mono-, or 1,1-disubstituted in order to prevent homopolymerization and uncontrolled autocatalytic reactions. If the vinyl monomer is 1,2-
10 substituted higher concentrations of about 10% by weight may be used.

As applicable vinyl monomers are mentioned crotonic acid, acrylic acid, acrylic acid esters, acrylamide, bisacrylamide, methylol acrylamide, acrylated amines, acrylated epoxides, acrylated ethers, acrylated polyester, acrylated polyurethanes, acrylated acrylates, acrylated silicones, acrylated silanes, acrylated phosphates and acrylated titanates, acrolein, phenyl substituted styrene derivatives as p-aminostyrene, tiglic
15 acid, seneciolic acid, angelic acid and cinnamic acid.

This process is difficult to control due to the inherent risk of excessive polymerization of the vinyl monomer. This is the most probable reason for the high binding capacity described. 5-7 µg protein per well can probably not be immobilized as a monolayer in one single micro-titre-well. It is therefore essential that the vinyl monomer is present in a chain transfer reducing solvent defined as a solvent which when irradiated
20 forms radicals not being able to initiate polymerization. As examples of suitable solvents are mentioned methanol, pyridine, water and mixtures of methanol and water. In practice 1:1 methanol/water mixtures are used.

Thus the known process is based on the recognition that vinyl monomers which are known to be polymerizable on polymer surfaces by irradiation graft polymerization or free radical graft polymerization
25 may be irradiation treated under conditions which prevent a polymerization and bound to the polymer surfaces as a thin graft layer close to a monomolecular layer leaving reactive groups capable of binding with biologically active molecules.

However, the known method suffer from a number of drawbacks. Firstly, the grafting process is very time-consuming. Even with γ-radiation which is the only tested radiation source, the reaction time is 10-12
30 hours. γ-radiation poses severe health physical requirements and there is a tendency of discolouring the plastic rendering it opal and therefore inapplicable for optical measurements.

Further the process requires an oxygen free atmosphere and a free radical initiator, e.g. benzophenone.

The documentation provided in the examples does not convincingly show that immobilization of proteins has been accomplished due to covalent binding to the grafted polymer. Further, a standard
35 coupling agent like glutaraldehyde or a carbodiimide reagent is used in most of the examples. These agents generally bring about a crosslinking of the protein molecules leading to an enhanced binding irrespective of the surface character.

Also, γ-radiation is known to activate polymer surfaces and thereby improve their protein binding properties.

40 In conclusion it has not been inambiguously substantiated that the reported protein binding results are due to an activation of the polymer surface and not a result of the γ-radiation and the use of coupling agents.

It has previously been described that bifunctional reagents containing arylazides can be bound to polymer surfaces (DE A 34 35 744). This was exemplified by adding of protein A to a solid phase and
45 subsequently adding the bifunctional compound: N-succinimidyl-6-(4'-azido-2'-nitrophenyl-amino)-hexanoate. This results in binding of the arylazide derivative to the amino groups of protein A. This conjugate was subsequently exposed to light and a covalent binding to the polymer solid phase was postulated. However, there is no evidence for a covalent binding between the polymer surface and protein A, since no data in the above-mentioned patent application showed the ability of protein A binding to the polymer
50 surface without addition of the arylazide compound.

Furthermore, during photolysis it is to be expected that degradation products of the azido group more likely will bind to the numerous nucleophilic groups present on protein A, thus forming aggregates of this protein. Furthermore, arylazides are known to react with nucleophiles as e.g. water, thereby further reducing the likelihood for a possible reaction with the polymer surface.

55 Investigations have been carried out in order to biotinylate polystyrene photochemically using the commercial available reagent N-(4-azido-2-nitrophenyl)-N'-(3-biotinylamino-propyl)-N'-methyl-1,3-propanediamine, (photobiotin, Sigma Cat. No. A 7667). This reagent contains an aryl azide group connected to a similar chemical linker as the one used according to EXAMPLE 1. The same optimizing experiments were

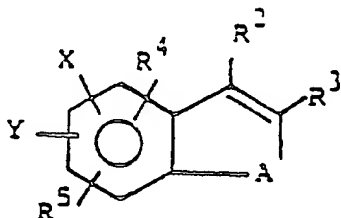
performed as in this example, but no biotinylation of the polystyrene solid phase could be detected.

DETAILED DESCRIPTION OF THE INVENTION

To obviate disadvantages associated with the modification methods presently existing, the present invention provides a method for modifying the surface of a polymer, especially a method for photo-chemical immobilization of molecules to polystyrene or other plastic materials.

The invention is based on the surprising finding that a number of well-known photoreactive polycyclic compounds are firmly bound to a variety of polymer surfaces if they are exposed to electromagnetic radiation under suitable conditions.

Accordingly, its broadest aspect, the invention relates to a method for modifying the surface of a polymer comprising contacting the polymer surface with a compound of the general formula I.



in which A is -O-, -S-, -Se-, >NH, >NR¹, -NH-O-, -N=N-, >S⁺R¹, -S-O-, >Se⁺R¹, -CO-O-, -CO-S-, -CS-O-, CS-S-, -CSe-O-, -CO-Se-, -CS-NH-, -CO-NH-, -CO-N-(R¹)-, >P=O or -P(=O)(O⁻)-;

R¹ is hydrocarbyl or hydrocarbyloxy, any of which may be substituted with NO, NO₂, SO₃, CN, OH, =O, SH, SeH, PO₃⁻⁻⁻, PO₂⁻, COO⁻, halogen epoxide, NH₂, NHR¹, NR¹R¹ (in which R¹ has the same meaning as defined for R¹).

R², R³, R⁴, and R⁵, which may be the same or different are H, halogen, NO, NO₂, SO₃, CN, OH, =O, SH, SeH, PO₃⁻⁻⁻, PO₂⁻, COO⁻, epoxide, NH₂, NHR¹, -NR¹R¹ or heterocyclyl, or has the same meaning as defined for R¹.

X and Y, which may be the same or different, have the same meaning as defined for R²-R⁵, or

X and Y are adjacent to one another and together form a group with the formula -CR⁶=CR⁷-A' where A' has the same meaning as defined for A above, and R⁶ and R⁷ which may be the same or different have the same meaning as defined for R²-R⁵; and

optionally, one or more of R¹-R⁷ designates a probe;

and irradiating the polymer and the compound of the formula I with electromagnetic radiation.

The suitable electromagnetic radiation may be chosen with respect to the photoreactive properties of the compound I, e.g. absorption-properties depending on the actual substituents and their surroundings, e.g. solvent and other solutes.

The electromagnetic irradiation is preferably performed at a wavelength shorter than 700 nm.

Presently preferred is irradiation in the UV-region, i.e. 10-400 nm. Depending of the actual compound I "long-wave" radiation (>350 nm) is preferred, but also "short-wave" (<350 nm) may be used.

As shown below, it is sufficient to apply incoherent UV-lamps, although application of more complicated light sources, e.g. monochromatic, non-monochromatic, polarized, non-polarized, coherent, non-coherent, continuous or discontinuous light-sources is contemplated.

The term "hydrocarbyl" designates groups comprising hydrogen and carbon such as alkyl, alkenyl and alkynyl groups, all comprising at the most 30 carbon atoms; aryl, alkaryl and aralkyl groups in which the "alk" or "alkyl" moieties comprise at the most 30 carbon atoms; and the term "aryl" designates phenyl, naphthyl, biphenyl, tolyl groups, etc.

The term "hydrocarbyloxy" designates a hydrocarbyl group as defined above connected to the remaining molecule via an oxygen bridge.

Examples of C₁₋₃₀ alkyl groups are methyl, ethyl, propyl, n-butyl, isobutyl, tert.butyl, n-, iso- and tert.pentyl, hexyl, heptyl, octyl, nonyl, decyl, dodecyl, pentadecyl, eiccsyl, pentacosyl, and triacontyl;

examples of C₂₋₃₀ alkenyl groups are ethenyl, propenyl-1, propenyl-2, buten-1-yl, buten-2-yl, pent-1-en-yl, hept-1-en-yl, etc.;

examples of C₂₋₃₀ alkynyl groups are ethynyl, propynyl, butynyl, pentynyl, etc.; and

examples of hydrocarbyloxy groups are hydrocarbyl groups as defined above, connected via an oxygen

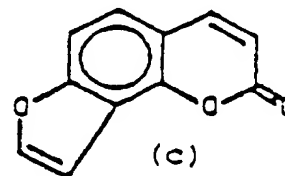
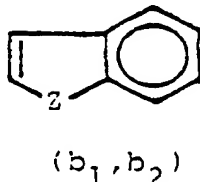
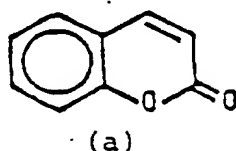
atom.

Especially preferred hydrocarbyl groups are long-chained alkyl groups, i.e. groups of 8-16 carbon atoms, especially methyl, undecyl, dodecyl, tridecyl, tetradecyl, pentadecyl, and hexadecyl.

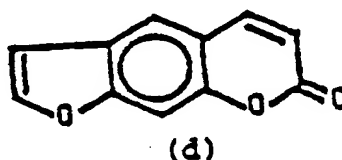
The term "heterocyclyl" preferably designates pyridyl, pyrimidinyl, pyridinyl, furyl, thienyl, imidazolyl, isoxazolyl, oxazolyl, thiazolyl, acridinyl and morpholyl, etc., which may be bound to the compound I in any position.

The term "halogen" designates fluoro, chloro, bromo, and iodo.

In an especially preferred embodiment of the invention, the compound I is selected from the group consisting of optionally substituted coumarins (a), benzofurans (b₁ with Z being O), indols (b₂ with z being NH) and angelicins (c)



The optionally substituted coumarins are preferably optionally substituted psoralens (d)



in which formulae (a)-(d) the optional substituents are as defined above for R²-R⁵.

In the following description reference is made to psoralens, but it is contemplated that other compounds encompassed by the general formula I can be utilized in the same manner as exemplified below.

The psoralens may comprise one or more substituents, and it is preferred that at least one of the substituents is an amino group containing constituent, preferably 3-tri-methylamino propoxy.

As example of optionally substituted psoralens may be mentioned psoralen, 8-methyl-3-carboxypsoralen, 4,5',8-trimethylpsoralen, 3'-trimethylamino-8-propyloxypsoralen.

Psoralens, which are a class of planar furocoumarin molecules capable of intercalating into double stranded deoxy-ribonucleic acid (dsDNA), will covalently bind to and crosslink DNA when activated by "long-wave" ($\lambda > 350$ nm) UV-light. The photoreaction takes place between thymidine and the double-bond in the coumarin and the furan part of psoralen under formation of a cyclic product. Coumarins are also known to be photo-reactive compounds (e.g. 5,7-dimethoxy coumarin) (Queval et al., Eur. J. Med. Chem. 9, 1974, p. 335).

According to the invention, psoralens can be bound to polymers such as polystyrene by means of electromagnetic irradiation and it is possible this way to immobilize molecules covalently to the polymers via psoralen.

The optionally substituted coumarins may further be exemplified by warfarin and the optionally substituted indols by 5-hydroxytryptamine and 3-indolacetic acid.

Polymers to be used in the method according to the present invention are preferably olefinic and can be selected from the group consisting of polystyrene, polyethylene terephthalate glycol, polyvinyl acetate, polyvinyl chloride, polyvinyl pyrrolidone, polyacrylo nitrile, polymethyl methacrylate, polytetrafluoro ethylene, butyl rubber, styrenebutadiene rubber, natural rubber, polyethylene poly-4-methylpentylene, polyester, and polypropylene. It is contemplated that also silica-containing materials such as glass can be used as a "polymer".

With the method according to the present invention, a non-premodified polymer is used. The method may be carried out in pure aqueous solution without addition of organic solvents. A very reproducible and uniform modification of the polymer is obtained and a transparent polymer such as polystyrene remains fully transparent during the whole procedure. It takes less than 60 min. to bind e.g. substituted psoralens in

this way depending on the light source and it is very easy to perform in large scale and involves no toxic or carcinogenic reagents.

As will appear from the following examples the efficiency of the polymer surface modifications also depends on the pH of the compound I solution. The optimal pH for a given compound i.a. depends on the individual substituents and their positive or negative charges and the stability of the compound and may be established by experiments. As a guideline, it is desirable to avoid protonization of compound I. Consequently alkaline pH is preferred for compounds having basic character, e.g. containing amino groups and an acidic pH is preferred for compounds having acidic character, e.g. containing carboxylic groups. For practical purposes a pH from about 5 to about 9 is preferred.

The efficiency of the polymer surface modification may be enhanced by irradiating the polymer and compound I in the presence of activator agents.

Preferably the present invention is performed in aqueous solution with optional presence of an ionic strength enhancing compound, e.g. NaCl.

Further, addition of benzoquinone to the solution surprisingly improves the modification of the polymer. The reason for this activator effect is not clear but either benzoquinone functions as a sensitizer or it participates directly in the binding of the compound I to the polymer surface.

An especially preferred embodiment of the present invention provides a method for linking a compound of the general formula II



in which L designates a linking unit (spacer arm) or a bond, and U designates a probe as further defined below

to a polymer surface via a compound of the general formula I as defined above.

The linking unit L is an organic moiety (spacer arm) or a bond capable of interconnecting the compound I and U. Examples of covalent or non-covalent linking units (spacer arms) are N,N'-dimethylcysteamine (Elsner et al., Anal Biochem. 149, 1985, p. 575-581), diamines such as N,N'-dimethyl hexane diamine, cysteamine, spermidine, norspermidine (Burchardt et al., JAGS 49, 1984, p. 4123-4126), piperazine (Hansen et al., Med Chem., 36, 1983, p. 1510-1514), seleno-dipropionic acid, glycerol, hexane, and diethyl ethene, and spacer arms as mentioned in EP A 156 287.

An example of compound I with attached linking unit is 8-propyloxypsoralenyl-N,N'-dimethyl hexane diamine, example of compound I, linker and probe is N'-biotinyl-N-8-propyloxypsoralenyl-N,N'-dimethyl hexane diamine.

The probe U is a chemical constituent of any kind which can be coupled covalently or non-covalently to L, thereby being immobilized to the polymer through the compound I. Examples of U are:

a) A label moiety which is capable of being identified using an appropriate technique. Examples of such techniques include spectroscopic, radioisotopic, photochemical, chemical, immunochemical or biochemical means as by using a polypeptide, lectin or antibody capable of forming a complex with the label moiety. Examples of label moieties are biotin, radioactive compounds, spin labels, fluorogenic compounds (e.g. fluorescein), enzymes or enzyme colourigenic substrate, pH colourigenic compounds (e.g. phenolphthaleine) or haptens which can be recognized using an e.g. enzyme labelled antibody. Thus, as used herein the term "label" is intended to include both moieties that may be detected directly and reactive moieties that are detected indirectly via a reaction which forms a detectable product.

b) Compounds comprising reactive chemical groups (e.g. the active esters N-hydroxy-succinimidooester and p-nitrophenylester) capable of reacting under the formation of covalent bonds with other chemical groups (e.g. amino groups on proteins). Other examples are active halogen-containing compounds (e.g. benzoyl bromide) which react with both thiol and amino groups, disulphide-containing compounds, epoxides or optically active compounds (e.g. brucine).

c) Non-covalent reacting chemical groups which comprise charged or hydrophobic chemical groups with affinity for compounds carrying an opposite charged group or a hydrophobic group. Examples of charged chemical groups are $-NH_3^+$ and $-SO_3^-$ with affinity for a compound carrying the opposite net charge (e.g. a protein).

d) Other molecules such as proteins with specific affinity for other compounds (e.g. enzymes, antibodies, lectins, hemoglobin, histones or non-histone proteins or hormones), peptides (e.g. peptide hormones), pharmaceuticals (e.g. cytostatics, vitamins or penicillin, nucleic acids (e.g. DNA probes), mono-, oligo-, or polysaccharides, and optionally substituted lipids.

Hence, the method according to the invention involves premodification of the molecules to be immobilized and not necessarily the polymer. This results in a very homogeneous modification of polystyrene and not in a heterogeneous mixture of functional groups as described above.

A further advantage of the method according to the invention in contrast to passive adsorption to plastic, is that it is possible to bind e.g. proteins to a solid phase (the polymer) despite of the presence of detergents in the coating solutions. This is often the case when detergent solubilized proteins are to be investigated.

5 Psoralens are furthermore known to photoreact with electrophils as e.g. double bonds. Arylazides are, on the other hand, known to photoreact with e.g. water and other solvents. This makes arylazides less suitable than psoralens.

All these advantages compared to the above-mentioned methods, implicate that the principle disclosed herein could be used for chemical solid phase synthesis as e.g. peptide- or oligonucleotide synthesis and a
10 special advantage is that the process can be followed spectrophotometrically directly in tubes or micro-titre-plates when the polymer is transparent.

Even if chemical synthesis in micro-titre-plates or tubes are not desired, the method according to the invention may as well be unsuitable for modification of polystyrene in another form as e.g. particles.

An interesting aspect of the invention is that it is possible to perform the process of binding substances
15 to polymers in arbitrary order. It is possible to photoreact the polymer and the compound I in the first step and then connect the linking unit (if necessary) and the probe. Another way is to build up a complex consisting of the compound I, the optional linking unit and the probe and then bind the complex formed to the polymer by photoactivation. Still another way is to start with a combination of a compound I and a linking unit, photoreact this combination with the polymer and finally add the probe.

20 In this connection it should be noted that the probe itself and/or the linking unit can constitute a part of the compound of the general formula I.

In general, a molecule can be immobilized on a solid surface by a method comprising the steps of

a) Reacting the molecule with a compound of the general formula I, optionally by means of a linking unit, contacting the combination with a polymer material and subjecting the polymer and combined
25 molecule and compound I to electromagnetic irradiation or,

b) modifying the surface of a polymer by reacting it with a compound I by means of electromagnetic irradiation and thereafter contacting the modified polymer surface with the molecule, optionally by means of a linking unit.

30 The method according to the present invention is very suitable for covalent immobilization of compounds (e.g. antigen or haptens) which do not passively adhere to plastics. The method according to the invention can be performed directly in micro-titre-plates, tubes, strips or on particles. Therefore, the method can also be utilized in the preparation of a solid phase to be used in different types of immunoassays.

In a special embodiment of the invention the solid surface is selected from a polymeric plate such as a
35 thin layer plate or micro-titre-plate, polymeric beads, strips, dipsticks, test tubes, and (micro)spheres.

In solid phase enzyme processing and affinity chromatography, proteins or other compounds with specific affinity to other molecules must be immobilized on a solid phase. A solution of the molecules to be separated or processed is then either passed through a column or added to a suspension containing the solid support. Degradation products or solid phase bound compounds can then be eluted subsequently.

40 Materials used as solid phases in chromatography are frequently based on agarose, polyacrylamide, dextran or cellulose. All these substances will collapse under the high pressures used in high performance liquid chromatography-(HPLC) techniques. Polystyrene microparticles covalently modified according to the invention is an advantageous alternative to the above-mentioned substances or silicon-containing materials used in HPLC-columns or other chromatographic systems.

45 In enzyme processing, solid phase immobilization is often desired. The method according to the invention could be used for this purpose as well as for immobilization of microbial cells or other cells. It is contemplated that immobilized enzymes could be used in purification of industrial waste and separating and inorganic compounds.

50 Fixation of e.g. enzymes to e.g. glass is used in biosensors. The present invention could be applied in the future development of these.

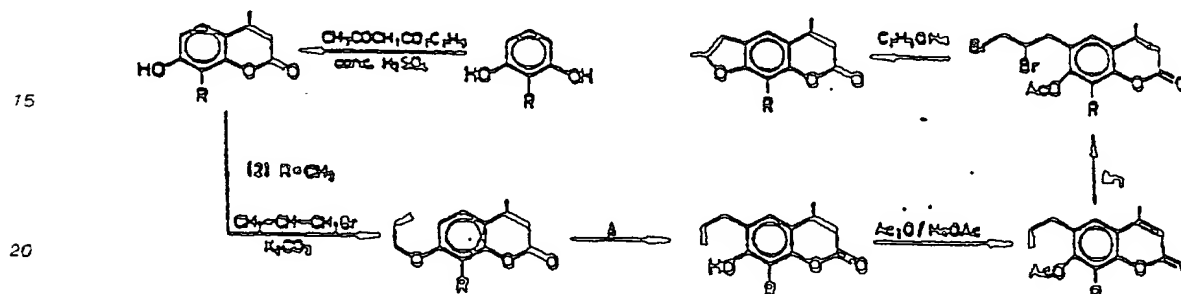
Microspheres have been used in the treatment of cancer using cytotoxic drugs immobilized to polymers. Furthermore, monoclonal anticancer cell antibodies immobilized on magnetic polystyrene particles have been used to separate cancer cells from bone marrow cells prior to autotransplantation. The present invention has application in these fields, too.

55 The present invention may even have application in the production of laminated plastic using e.g. psoralen compounds as photo-active glue between the laminae. It may also be used for binding coloured compounds covalently to e.g. polystyrene using e.g. psoralens conjugated to a coloured compound.

Compounds I can be synthesized according to the following principles:

Photo-active benzofurans, coumarins as psoralens and angelicins can be synthesized or they can be isolated from plants or coal tar. 4,5',8-Trimethylpsoralen and 8-methoxypsoralen can be isolated from the fungus *Sclerotinia sclerotiorum* (Scheel et al., Biochem. 2, 1963, p. 1127), and benzofurans and coumarins can be isolated from coal tar or plants.

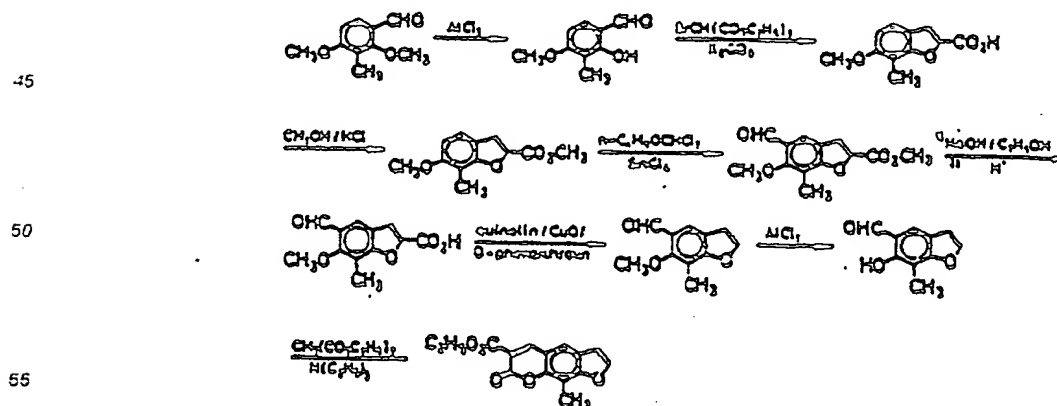
- 5 Psoralens, e.g. 4,5',8-substituted psoralens, can be synthesized from 2-methyl-3-hydroxy-phenol by reaction with acetoacetic acid ethyl ester, which leads to formation of a substituted coumarin (Rangaswani et al., Proc. Ind. Acad. Sci. Sect. A6, 1937, p. 112). After 5 further synthesis steps: reaction with 3-bromo-2-ene propane, heating, treatment with base, reaction with bromine and finally treatment with sodium ethanolate, a 4,5',8-substituted psoralen is formed (Queval et al., Eur. J. Med. Chem. 9, 1974, p.335). The
10 reaction sequence is illustrated below:



- 25 By replacing 2-methyl-3-hydroxy-phenol with other substituted phenols, it is possible to replace the substituents on the coumarin product and consequently the 4,5',8-substituted psoralen of the formula I. Using 2-methoxy-3-hydroxy-phenol instead, the end product will be 4,5'-dimethyl-8-methoxypsoralen instead of 4,5'-dimethyl-8-methylpsoralen. If the ester group in the coumarin is to be replaced by e.g. an amide, thioester, phosphoester or selenoester, other phenol reagents can be used as starting material, for example a starting material in which the 3-hydroxy group in the hydroxy phenol is replaced by an amine,
30 thiol, phosphor or seleno. If the starting material was 2-amino-3-methyl-phenol, the product would have been a coumarin with an amide instead of an ester (lactam instead of lactone).

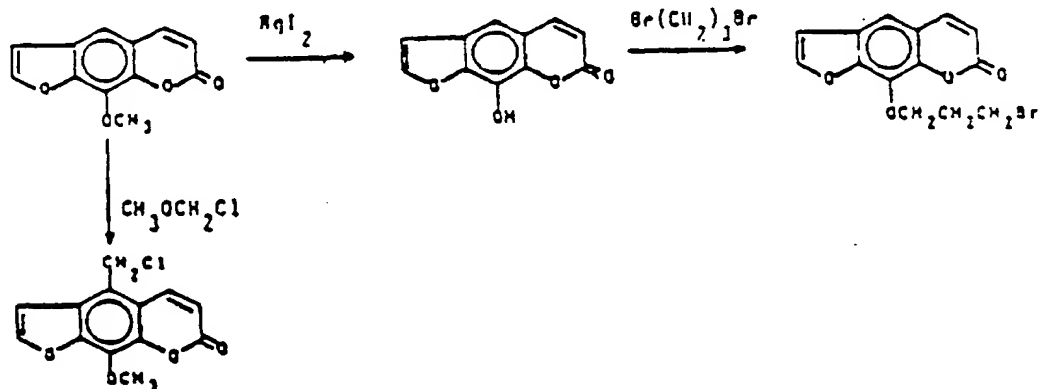
- By replacing the acetic acid ethyl ester with a haloalkanone in the first step of the synthesis, the ester group in the coumarin can be further substituted. If the acetoacetic acid ethyl ester is replaced by 1-bromo-3-butanone, $(\text{CH}_3\text{COCH}_2\text{CH}_2\text{Br})$, when reacting with 2-methyl-3-hydroxy-phenol or 2-amino-3-hydroxy-phenol, the product will be a coumarin analogue in which the ester is replacing an ether and an amine,
35 respectively.

- Synthesis of psoralens can also be carried out by initially synthesizing the benzofuran part. It is possible to start from 2,4-dimethoxy-3-methyl benzaldehyde and reduce this compound with aluminium trichloride and react with diethyl bromomalonate. This leads to substituted benzofurans (Queval et al., Eur. J. Med. Chem. 9, 1974, p. 335). Further reactions in six steps lead to 3-carbomethoxy-8-methylpsoralen. The
40 reaction sequence is illustrated below:



If the carbethoxypsoralen is treated with glacial acetic acid in sulphuric acid, a psoralen containing a carboxylic acid is produced.

Modifying the compound I with a reactive group can be performed with chloromethylmethyl ether resulting in a chloromethylation product depending on the type of compound I used. If the compound I is 8-methoxypsoralen, the product will be 5-chloromethyl-8-methoxypsoralen, and if the compound I is 4,5'-8-trimethylpsoralen, the product will be 4,5',8-trimethyl-5'-chloromethylpsoralen (Elsner et al., Anal. Biochem. 149, 1985, p. 575-581). It is also possible to convert a methoxy group into a reactive group by reacting it with magnesium iodide and dibromopropane. If the compound I used is 8-methoxypsoralen, the resulting product will be 3-bromopropoxy-8-methoxypsoralen. The conversions mentioned above are illustrated below:



A reactive ester group such as a N-hydroxy-succinimido ester can be introduced into a compound I containing a carboxylic group. The introduction can be performed by reacting the carboxylic group with N-hydroxy-succinimide in the presence of a carbodiimide in e.g. dioxane.

Introduction of a linking unit (spacer arm) (if necessary) can be performed using different synthetic routes. It seems to be possible to connect a linking unit at all positions on the compound I. To the linking unit any compound (e.g. biotin, enzyme, chiral compounds, pharma, etc.) can be connected. If 3-bromopropoxypsoralen is heated with N'-tert.butyloxycarbonyl-N,N'-dimethylhexane-diamine in acetone, the linking unit, N'-tert.butyloxycarbonyl-N,N'-dimethyl hexane diamine, will be bound covalently to the compound I. Thereafter, the tert.butyloxycarbonyl can be replaced with e.g. biotin. As examples of other groups could be mentioned an arylazide or other photoreactive compounds (Elsner et al., Anal. Biochem. 149, 1985, p. 575-581).

In the specification the following abbreviations are used:

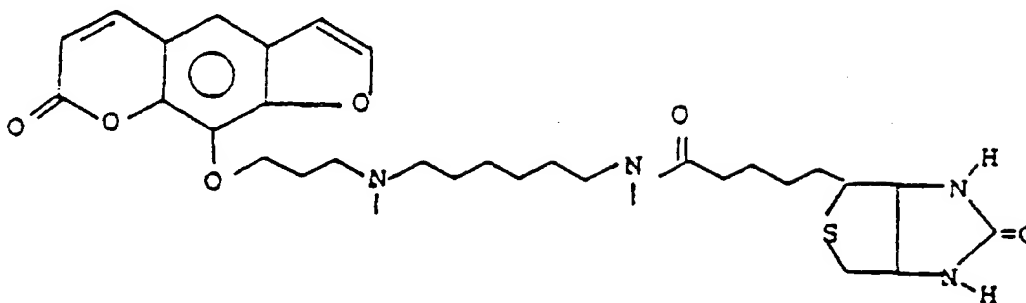
ELISA: enzyme linked immunosorbent assay
 dsDNA: double stranded deoxy-ribonucleic acid
 HPLC: high performance liquid chromatography
 HRP: horseradish peroxidase
 BSA: bovine serum albumine
 OD: optical density
 IgM: immunoglobulin of the M class
 IgG: immunoglobulin of the G class
 RF: rheumatoid factor
 DMF: dimethyl formamide
 DMSO: dimethyl sulphoxide
 PMMA: polymethacrylate particles
 Boc: tert.butoxycarbonyl
 PBS: phosphate buffer saline

The present invention will be further illustrated in Examples 1-11 below and binding of compounds II to psoralen will be illustrated in Examples A-C below:

EXAMPLE 1

Biotinylation of polystyrene with PS12

Biotin-conjugated psoralen (PS12) with the formula



was synthesized as follows:

Tert-butoxycarbonyl-N,N'-dimethyl hexane diamine (1)

Tert-butoxycarbonyl azide in ether ("Organic Synthesis V", p. 157-158) (2.9 g, 20 mmol, 5.2 ml) was added dropwise to a stirred solution of N,N'-dimethyl-hexane diamine (Aldrich Cat. No. D16110-1) (Beilstein 4(1), p. 422) (2.92 g, 20 mmol) in dimethyl sulphoxide (DMSO) (30 ml). The temperature was kept at 20 °C with external cooling. After the addition was complete (30 minutes), stirring was continued at room temperature for 1 hour, whereupon water (30 ml) was added. The solution was acidified (pH 5) with 2N HCl and extracted with ether (2 x 200 ml). The DMSO/water-mixture was made alkaline (pH 11) with 6N NaOH and extracted with ether (4 x 200 ml). Evaporation of the ether gave the title compound (1) (1.2 g) as a yellow oil.

8-Hydroxypsoralen (2)

To a suspension of magnesium (0.97 g, 0.04 mol) in dry benzene (75 ml) iodine (10.2g, 0.04 mol) was added over a period of 1 hour. Then 75 ml of dry benzene was added and the mixture was refluxed for 1 hour. Next day 8-methoxypsoralen (Sigma Cat. No. N 3501) (4.3 g, 0.02 mol) was added, and after stirring for 15 minutes the solvent was evaporated under vacuum at 120 °C until the residue was practically dry. It was then further heated at 165 °C for 2 hours. The resulting solid was decomposed with dilute sulphuric acid and filtered. The solid was washed with water, suspended in dilute bisulfite solution, filtered again, washed with water and finally crystallized from dioxane and yielded colourless crystals (3.75 g), m.p. 246 °C.

3-Bromo-8-propyloxypsoralen (3)

8-Hydroxypsoralen (2) (4.04 g, 20 mmol) and dibromopropane (1.6 g, 8 mmol) were refluxed in acetone (400 ml) with potassium carbonate (20 g) for 24 hours. The solution was filtered and evaporated in vacuum. Chromatography on the residue on silica gel with chloroform as eluent yielded 3-bromo-propyloxypsoralen (2.6 g) as a white powder.

PS12

3-Bromo-propyloxypsoralen (3) (1.6 g) and N-tert-butoxycarbonyl-N,N'-dimethyl + hexane diamine (1) (1.3 g) were mixed in acetone (100 ml) with potassium carbonate (1.8 g) and refluxed for 72 hours, whereupon it was filtered and evaporated in vacuum. The residue was stirred in HCl (2M) in glacial acetic acid for 1 hour and subsequently evaporated under vacuum. The residue was solubilized in absolute ethanol (20 ml) and triethyl amine (1 ml), whereupon hydroxysuccinimido-biotin (NHS-d-biotin), (Sigma Cat. No

H1759), (1.7 g) was added. After stirring for 24 hours it was evaporated in vacuum, and diluted with chloroform (2 x 2 ml), the solution was made alkaline with diluted sodium hydroxide and extracted again with chloroform (3 x 2 ml). The basic chloroform extracts were pooled, dried over magnesium sulphate and evaporated in vacuum. The residue was solubilized in 1 ml methanol, whereupon 100 μ l hydrochloric acid (10N) was added. The hydrochloride (PS12) was precipitated with ether, collected and freeze dried.

PS12-binding to polystyrene

A 10 mg/ml stock solution of PS12 in dimethyl sulphoxide was stored at -4°C . Ten-fold dilutions in distilled water were made from this stock solution in concentrations ranging from 0.1 to 1,000 $\mu\text{g/ml}$, and 100 $\mu\text{l/well}$ of each dilution were added to the wells of polystyrene micro-titre-plates (Nunc, Denmark). The plates were then irradiated for 2 hours at room temperature with "long-wave" ($\lambda > 350\text{nm}$) UV-light from a Philips TL 20W/09N lamp placed 20 cm above the micro-titre-plates, and the wells were washed 8 times with 200 μl washing buffer (29.2 g NaCl, 0.2 g KCl, 0.2 g KH_2PO_4 , 0.92 g Na_2HPO_4 , Triton®X-100, water ad 1,000 ml, pH 7.2). The same experiment was carried out without irradiation of micro-titre-plates.

For detection PS12-binding to the solid phase, a solution of 100 $\mu\text{l/well}$ of an avidin-horseradish peroxidase (avidin-HRP) complex (Sigma), 25 μg avidin-HRP, 100 mg BSA, 10 ml washing buffer was added to the wells and incubated at 37°C for 1 hour. The plates were then washed 3 times with washing buffer and 100 $\mu\text{l/well}$ of a colourigenic substrate were added (1 mg o-phenyl diamine (Sigma) per ml citrate/phosphate buffer (7.3 g citric acid, 11.86 g Na_2HPO_4 , H_2O_2 , distilled water ad 1 liter), 1 μl H_2O_2 - (35%). After approximately 10 minutes the colour reaction was stopped with 100 ml 1N H_2SO_4 and the optical density (OD) was read on a Titertek® Multiscan ELISA-photometer (Fig. 1).

Time dependency of psoralen-biotin binding

A PS12 stock solution was diluted in water to a concentration of 10 $\mu\text{g/ml}$ and 100 $\mu\text{g/ml}$, respectively. 100 $\mu\text{l/well}$ of each dilution were transferred to micro-titre-plates, UV-irradiated with different duration or irradiation time and treated as described above (Fig. 2).

Immobilization of streptavidin

Streptavidin (Zymed) (1 $\mu\text{g/well}$ in distilled water) was added to each well of micro-titre-plates coated with PS12 (0.1 $\mu\text{g/well}$, 1 $\mu\text{g/well}$ and 10 $\mu\text{g/well}$). After 12 hours at 4°C , the plates were washed 3 times with washing buffer, and a solution of 100 μl of a biotin-HRP complex (Sigma) (25 μg biotin-HRP, 100 mg BSA, 10 ml washing buffer, water) was added, after which the plates were incubated at 37°C . After 1 hour, the plates were washed 3 times with washing buffer and peroxidase activity was detected as described above (Fig 3).

RESULTS

It was shown that PS12, when irradiated with UV-light, was able to bind to polystyrene. PS12-binding was measured with avidin-HRP. Without UV-irradiation no biotinylation of the wells could be detected (Fig. 1). This strongly indicates that the binding is of covalent nature. This is further supported by the fact that PS12 cannot be removed by subsequent treatment under strong alkaline or acidic conditions or with most organic solvents. Thus the binding resists overnight treatment with e.g. 4N, H_2SO_4 , 10 N NaOH, abs. ethanol or methanol glacial acetic acid or 50% DMSO in water.

Biotinylation of polystyrene increased when increasing concentrations of PS12 were added to the wells. No maximum values were achieved in these experiments. OD-valued increased from 0.2 to 2.6 with PS12 concentrations ranging from 0.1 $\mu\text{g/well}$ to 1,000 $\mu\text{g/well}$.

Using avidin-HRP it was shown that a maximum biotinylation degree was reached after an irradiation period of 1.5 hours. OD-values increased from 0.1 to 1.2 during this period when 1 μg PS12 was used and from 0.1 to 1.6 when 10 μg PS12 was used (Fig. 2).

A maximum number of free biotin binding sites on immobilized streptavidin, as detected with biotin-HRP, was achieved when wells were coated with 1 $\mu\text{g}/100 \mu\text{l}$ PS12. This corresponded to an OD-value of

1.3. When the plates were coated with 0.1 $\mu\text{g}/\text{well}$ or 10 $\mu\text{g}/\text{well}$ PS12, the number of free biotin sites were less. The decrease in the number of biotin sites using PS12 concentrations less than 1 $\mu\text{g}/\text{ml}$ was probably due to low amounts of biotin on the polystyrene surface. Since it was shown in Fig. 1 that the amount of avidin-HRP immobilized to the biotin-conjugated polystyrene surface correlates well with PS12 concentrations, the decrease in free biotin sites seen in Fig. 3 can be explained as a blocking of the biotin sites on streptavidin by increasing amounts of solid phase bound biotin.

When no PS12 were used, no streptavidin was bound to polystyrene as it appears from Fig 3.

When 1 $\mu\text{g}/\text{well}$ PS12 was used, the biotinylation degree, as detected with avidin-HRP, gave an OD > 3, whereas no detectable free biotin sites were observed when using biotin-HRP (Table 1). When 2 $\mu\text{g}/\text{well}$ streptavidin were added to each well, the amount of free solid phase bound biotin, measured with avidin-peroxidase decreased to OD = 0.111. Concurrently, the amount of free biotin sites, measured with biotin-HRP, increased to OD > 3 (Table 1). This indicates that streptavidin actually was immobilized and bound to nearly all of the solid phase bound biotin. Using biotin- or avidin-conjugated peroxidase, it was furthermore possible to follow the subsequent binding of biotinylated compounds as e.g. nucleic acids.

TABLE 1

	PS12-coated wells	PS12- + streptavidincoated wells
Biotin-peroxidase	0.070	>3.000
Avidin-peroxidase	>3.000	0.111

EXAMPLE 2

An enzyme linked immunosorbent assay (ELISA) for determination of IgM rheumatoid factors

Passive adsorption of avidin to a solid phase has been described in connection with secondary immobilization of biotinylated compounds such as e.g. immunoglobulin or carbohydrates (US Patent No. 4,582,810). The exemplified use for this invention was development of immunoassays.

It is, however, well known that many healthy persons possess anti-avidin antibodies in their sera, which may disturb the measurement of other compounds.

Streptavidin, isolated from *Streptomyces* shows, on the other hand, very low reactivity with normal human serum, but do not, unlike avidin, adhere passively to polystyrene (EXAMPLE 1). Streptavidin therefore can not be used in this way as an agent for secondary immobilization of biotinylated compounds.

However, with the method according to the present invention, streptavidin can be immobilized to polystyrene. This can be used in e.g. enzyme, linked immunosorbent assays (ELISA). This is exemplified by an ELISA-method for determination of IgM-rheumatoid factors (IgM-RF).

These are antibodies of the IgM immunoglobulin class, which have affinity for the Fc (fragment crystallizable) part of immunoglobulin of the IgG class. This method can be used for diagnosis of rheumatoid arthritis.

Passive binding of avidin to polystyrene micro-titre-plates

Solutions (100 μl) of avidin in concentrations ranging from 1 ng/ml to 1 mg/ml in 0.1 M carbonat buffer, pH 9.5, were added to each well of non-modified polystyrene micro-titre-plates and incubated overnight at 4°C. The plated were washed with washing buffer and presence of biotin sites was detected with biotin-HRP as described in EXAMPLE 1 (Fig. 4).

Immobilization of streptavidin on polystyrene micro-titre-plates was performed as described in EXAMPLE 1. 1 μg PS12 was added to each well and the plates were irradiated for 2 hours at room temperature with UV light at $\lambda > 350$ nm. Subsequently 2 $\mu\text{g}/\text{well}$ of streptavidin were added, incubated for 1 hour at 37°C and washed 3 times as described in EXAMPLE 1. Other non-modified polystyrene micro-titre-plates

were coated directly with 2 µg/well avidin as described above.

The streptavidin- and avidin-coated plates were blocked overnight at 4 °C with washing buffer containing 1% BS (dilution buffer) (200 µl/well). One plate containing dilution buffer only was also prepared.

Reactivity of immunoglobulin from healthy blood donors with solid phase immobilized avidin and streptavidin, respectively.

Sera from 10 healthy blood donors were tested on plates coated with streptavidin, avidin and BSA, respectively, by adding 100 µl/well, 1:100 dilutions in dilution buffer and incubating for 1 hour at room temperature. After a 1 hour incubation period the plates were washed and a 1:1000 dilution of HRP-labelled rabbit-antihuman IgM or IgG was added (100 µl/well) and incubated 1 hour at room temperature. Solid phase bound HRP was detected as described in EXAMPLE 1.

Determination of IgM-RF using biotinylated bound IgG bound to solid phase immobilized streptavidin

Biotinylated human IgG (VECTOR) was immobilized to PS12-modified polystyrene micro-titre-plates via solid phase immobilized streptavidin.

A solution of 20 µl/ml biotinylated IgG in dilution buffer (100 µl/well) was added to the plates containing immobilized streptavidin as described above, and the plates were incubated for 1 hour at 37 °C. They were then washed and serum dilutions (1:100) in dilution buffer of 10 IgM-RF positive sera and sera from 10 healthy blood donors were added in duplicate (100 µl/well).

Binding of immunoglobulin of the IgM-class was detected as described above.

RESULTS

In Fig. 4 it is shown that the amount of passively adsorbed avidin increases when increasing amounts are added to the wells. Maximum OD-values are obtained when 2 µg are added to each well.

In EXAMPLE 1 it was shown that no streptavidin was adsorbed when using non-biotinylated polystyrene. This was confirmed using 125-I labelled streptavidin.

Fig. 5 shows the results of testing serum from 10 healthy blood donors on wells coated with avidin, streptavidin and BSA, respectively. Compared to BSA-coated wells, a significant number of sera possess IgG as well as IgM-antibodies directed against avidin. No significant reaction is, however, seen with streptavidin-coated wells.

Fig. 6 shows the results of testing sera from the same 10 donors and furthermore 10 IgM-RF positive sera for the presence of IgM-RF. This was done using micro-titre-plates containing streptavidin immobilized human biotinylated IgG. All IgM-RF positive sera were also found positive in this ELISA-assay and none of the sera from healthy blood donors contained IgM-RF.

In this way biotinylated IgG was bound very strongly to the solid phase which must be advantageous when the plates are stored in dry state.

EXAMPLE 3

Biotinylation of Polymethylmethacrylate with PS12

To 3 beakers with 2 ml 0.05 NaCl in water 100 mg polymethylmethacrylate particles (PMMA) ELAVASIT 2041 (Dupont) were added. To 2 of the beakers with PMMA were added 10 µl PS12 (10 mg/ml in DMSO (dimethyl sulphoxide)). One of the beakers was irradiated for 1 hour with UV-light ($\lambda > 350$ nm) from a Philips TL 20W/09N lamp 20 cm above the beaker and the content was stirred. After irradiation, the PMMA-particles were washed with 3 x 1 ml washing buffer (29.2 g NaCl, 0.2 g KCl, 0.2 g KH₂PO₄, 0.92 g Na₂HPO₄, Triton®X-100, water ad 1,000 ml, pH 7.2) and w times with water. Then a solution of 1 ml avidin-peroxidase (Sigma) (25 µg avidin-peroxidase, 100 mg BSA, 10 ml washing buffer) was added, and the beakers were incubated at 37 °C. After 1 hour the PMMA-particles were washed 3 times with washing buffer, transferred to clean Eppendorf tubes and 1 ml substrate was added (3 mg 2,2'-azinobis-(3-ethylbenzthiazoline sulphonic acid) (Sigma) in 10 ml water). After 5 minutes 100 µl citric acid (10 g in 100 ml water) were added to each beaker and the optical density (OD) at 410 nm was determined after dilution with water 10 times.

RESULTS:

TABLE 2

Polymethylmethacrylate:	0.030 OD
Polymethylmethacrylate + PS12:	0.251 OD
Polymethylmethacrylate + PS12 + light:	1.010 OD

CONCLUSION

PS12 binds photochemically to PMMA.

EXAMPLE 4

Biotinylation of polystyrene with PS13

Aqueous solutions of radioactive labelled ^3H -3-trimethylamine-8-propoxypsoralen (PS13) (J.B. Hansen et al., Journal of Medical Chemistry 28, 1985, p. 1001-1010) were added to polystyrene micro-titre-wells (100 μl , 1 mg/ml, 3,800,000 cpm) (NUNC, Denmark) and diluted ten-fold 6 times with distilled water to a final concentration of 0.01 $\mu\text{g/ml}$. The micro-titre-wells were then irradiated as described above for 2 hours at room temperature and washed 8 times with 200 μl washing buffer. The same experiment were performed without UV-irradiation.

The wells were then emptied, separated mechanically and transferred to scintillator vials where solid phase immobilized radioactivity was determined using liquid scintillation counting in 10 ml Instagel (Packard).

RESULTS

At 1 mg/ml 87 ng PS13 were photochemically immobilized whereas only 17 ng were immobilized in non-irradiated wells. When using a concentration of PS13 at 100 $\mu\text{g/ml}$, 48 ng were immobilized to the solid phase whereas non-irradiated wells showed values near background values (12 μg) (Fig. 7).

At concentrations less than 100 $\mu\text{g/ml}$, no significant immobilization of PS13 could be detected - perhaps due to the low level of radioactivity in the dilutions compared to background values.

EXAMPLE 5

Biotinylation of micro-titre-plates with PS12

Similar experiments as described in EXAMPLE 1 using 2 μg PS12 per well were performed using micro-titre-plates of difficult manufacture and materials.

The following flat bottom micro-titre plates were used:

Titertek®:	Polyvinyl chloride, Cat. No. 77-173-05, Flow Laboratories, USA (FlowPVC);
Nunc-Immunoplates:	Polystyrene, Cat. No. 439454, Nunc, Denmark (NuncPS);
Polystyrene with high binding capacity:	Cat. No. 655061, C.A. Greiner und Söhne, West-Germany, (GreinPSH);
Polystyrene with medium binding capacity:	Cat. No. 655001, C.A. Greiner und Söhne, West-Germany, (GreinPSL);
Polystyrene EIA-plates:	Cat. No. 3590, Costar, USA (CostPS);
Polyethylene glycol terephthalate plates with high binding capacity:	Cat. No. 6595, Costar, USA, (CostPETG).

RESULTS

As shown in Table 3, all plate types bound large amounts of PS12 when irradiated with UV-light as defined in EXAMPLE 1. Non-irradiated plates bound no significant amounts of PS12.

TABLE 3

Plate type	Irradiated	Non-irradiated
FlowPVC	1.09	0.20
NuncPS	1.11	0.12
GreinPSH	0.90	0.10
GreinPSL	1.45	0.08
CostPS	0.84	0.10
CostPETG	0.68	0.11

EXAMPLE 6

Influence of ionic strength on biotinylation of micro-titre-plates

Similar experiments were performed as described in Example 5 using NaCl-solution (1M) instead of distilled water during the irradiation period. The biotinylation degree of the polymers was increased by 50-100%

EXAMPLE 7

Immobilization of 5-hydroxytryptamine to polystyrene

A solution of 10 mg/ml 5-hydroxytryptamine (Aldrich catalog number 10775-1) in DMSO was prepared. Respectively 0.1, 1 and 10 μ l of the 5-hydroxytryptamine in DMSO was added to a carbonate buffer (pH=5, 3 x 1 ml, 0.1 M), a phosphate buffer (pH=7, 3 x 1 ml, 0.1 M), and a citrate buffer (pH=9, 3 x 1 ml, 0.1 M). To each of the 9 solutions were then added 3 H-labelled 5-hydroxytryptamine (Amersham catalog number TRK.223) (10 μ Ci, 10 μ l, 4 μ g) which resulted in 9 different solutions with different pH and concentrations. The pH is 5, 7 and 9 and the concentrations of 5-hydroxytryptamine was 5, 14 and 104 μ g/ml buffer.

To 2 x 9 polystyrene micro-titre-wells (Cova-sorb, NUNC Denmark) was added 2 x 100 μ l buffer with 5-hydroxytryptamine from each of the 9 solutions. The micro-titre-wells were then irradiated with "long-wave"

($\lambda > 350$ nm) UV-radiation from a Philips lamp TL 20W/09N for 3 h at room temperature and washed 8 times with 200 μ l washing-buffer and 2 times with 100 μ l ethanol as described above in EXAMPLE 1.

The same experiments were performed without UV-irradiation, and with short-UV radiation from a Philips lamp 57415-P'40 TUV 15 W. The wells were then emptied, separated mechanically and transferred to scintillator vials, where solid phase immobilized radioactivity was determined using liquid scintillation counting, in 1.5 ml Ecoscient A (National Diagnostic).

The amount of immobilized 5-hydroxytryptamine is determined from a standard curve prepared from a 10 fold dilution of buffer with 5-hydroxytryptamine (14 μ g/ml).

RESULTS

The amount of immobilized 5-hydroxytryptamine depended of the pH and type of UV-light (TABLE 4). The amount of immobilized 5-hydroxytryptamine was largest after irradiation with short UV-light at neutral pH, i.e. 8 ng/well.

TABLE 4

pH	Conc μ g/ml	Dark*	Radiation long UV*	Radiation short UV*
9	5	1	20	300
9	14	10	120	700
9	104	0	120	80
7	5	1	3	310
7	14	80	70	5000
7	104	500	900	8000
5	5	0	12	0
5	14	10	100	90
5	104	1100	1000	3000

*pg 5-hydroxytryptamine immobilized.

EXAMPLE 8

Immobilization of 3-indolacetic acid to polystyrene

A solution of 10 mg/ml 3-indolacetic acid (Aldrich catalog number I-375-0) in DMSO was prepared. Respectively 0.1, 1 and 10 μ l of the 3-indolacetic acid in DMSO was added to a carbonate buffer (pH=9, 3 x 1 ml, 0.1 M), a phosphate buffer (pH=7, 3 x 1 ml, 0.1 M), and a citrate buffer (pH=5, 3 x 1 ml, 0.1 M). To each of the 9 solutions was then added 14 C-labelled 3-indolacetic acid (Amersham catalog number CFA-323) (1 μ Ci, 20 μ l, 3 μ g) which resulted in 9 different solutions with different pH and concentrations. The pH was 5, 7 and 9 and the concentrations of 3-indolacetic acid is 5, 14 and 104 μ g/ml buffer.

To 2 x 9 polystyrene micro-titre-wells (Cova-sorb, NUNC Denmark) was added 2 x 100 μ l buffer with 3-indolacetic acid from each of the 9 solutions. The micro-titre-wells were then irradiated with "long-wave" ($\lambda > 350$ nm) UV-radiation from a Philips lamp TL 20W/09N for 3 h at room temperature and washed 8 times with 200 μ l washing-buffer and 2 times with 100 μ l ethanol, as described above in example 1. The same experiments were performed without UV-irradiation, and with short-UV radiation from a Philips lamp 57415-P'40 TUV 15 W. The wells were then emptied, separated mechanically and transferred to scintillator vials, where solid phase immobilized radioactivity was determined using liquid scintillation counting, in 1.5 ml Ecoscient A (National Diagnostic).

The amount of immobilized 3-indolacetic acid was determined from a standard curve that was prepared from a 10 fold dilution of buffer with 3-indolacetic acid (13 μ g/ml).

RESULTS

The amount of immobilized 3-indolacetic acid depended of the pH and type of UV-light (TABLE 5). The amount of immobilized 3-indolacetic acid was largest after irradiation with short UV-light, i.e. 12 ng.vials.
 5 With long UV light it is possible to immobilize 11 ng/well at pH=5.

TABLE 5

pH	Conc μg/ml	Dark*	Radiation long UV*	Radiaton short UV*
9	4	7	6	7
9	13	70	30	70
9	103	230	200	400
7	4	2	8	70
7	13	100	30	30
7	103	800	1000	1200
5	4	1	12	80
5	13	7	100	100
5	103	0	1100	9000

*pg 3-indolacetic acid immobilized.

EXAMPLE 9

Immobilization of warfarin to polystyrene

A solution of 10 mg/ml warfarin (Aldrich catalog number 10775-1) in DMSO was prepared. Respectively 0.1, 1 and 10 μl of the warfarin in DMSO was added to a carbonate buffer (pH=9, 3 x 1 ml, 0.1 M), a phosphate buffer (pH=7, 3 x 1 ml, 0.1 M), and a citrate buffer (pH=5, 3 x 1 ml, 0.1 M). To each of the 9 solutions was then added ³C-labelled warfarin (Amersham catalog number CFA.449) (1 μCi, 20 μl, 7 μg) which results in 9 different solutions with different pH and concentrations. The pH is 5, 7 and 9 and the concentrations of warfarin is 8, 17 and 107 μg/ml buffer.

To 2 x 9 polystyrene micro-titre-wells (Cova-sorb, NUNC Denmark) was added 2 x 100 μl buffer with warfarin from each of the 9 solutions. The micro-titre-wells were then irradiated with "long-wave" UV-radiation from a Philips lamp TL 20W/09N for 3 h at room temperature and washed 8 times with 200 μl washing-buffer and 2 times with 100 μl ethanol as described above in EXAMPLE 1.

The same experiments were performed without UV-irradiation, and with short-UV radiation from a Philips lamp 57415-P/40 TUV 15 W. The wells were then emptied, separated mechanically and transferred to scintillator vials, where solid phase immobilized radioactivity was determinated using liquid scintillation counting, in 1,5 ml Ecoscient A (National Diagnostic).

The amount of immobilized warfarin was determinated from a standard curve that was prepared from a 10 fold dilution of buffer with warfarin (17 μg/ml).

RESULTS

The amount of immobilized warfarin depends of the pH and type of UV light. The amount of immobilized warfarin was largest after irradiation with long UV-light (TABLE 6), 300 ng/well at ph = 5.

TABLE 6

pH	Conc μg/ml	Dark*	Radiation long UV*	Radiation short UV*
9	8	0	808	7
9	17	10	20	20
9	107	200	200	80
7	8	0	1	5
7	17	0	20	20
7	107	60	90	80
5	8	1	2	10
5	17	0	20	100
5	107	0	300	200

*pg warfarin immobilized.

20 EXAMPLE 10

Enhanced coupling of PS12 onto polystyrene MicroWells using activator molecules, e.g. para-benzoquinone

25 A stock solution of para-benzoquinone, Merck Art. 802410, was made with a concentration of 10 mg/ml DMSO. The stock solution was stored at +4 °C.

PS12 (stock solution 10 mg/ml DMSO) was diluted to 4 μg/ml in phosphate buffered saline, pH 7.3, 2 M CaCl. Para-benzoquinone was added in a 2-fold dilution series (to give final concentrations from 0-500 μg/ml).

30 100 μl of the PS12 para-benzoquinone suspension was pipetted to each well of a MicroWell module (Nunc-Immuno Module C12, A/S NUNC, Denmark).

The coupling was done by irradiating the filled wells for 1 hour using a Philips UV lamp, type TL20W/09N. The distance between the lamp and the MicroWell was 20 cm.

Each well was washed 3 times using 350 μl washing solution (phosphate buffered saline, pH 7.2, 2 M NaCl, Triton®X-100), and the coupling of PS12 onto the MicroWells was detected by adding a mixture of 35 avidin (Sigma No. A-9390) and horse radish peroxidase conjugated to avidin (Dakopatts code P347, lot 047), 100 μl/well, at a concentration of 4 μg/ml and 0.45 μg/ml, respectively.

Incubation for 2 hours at room temperature, and washing 3 times with washing buffer (350 μl/well).

Substrate reaction was made according to the procedure described in EXAMPLE 1.

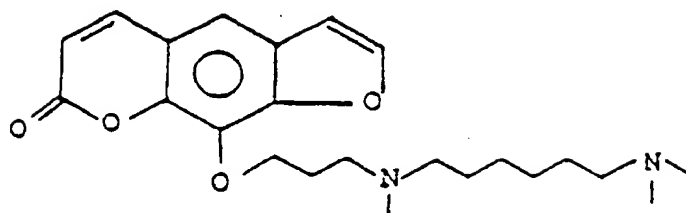
40 The optical density was read using a Jasco Immuno Reader NJ-2000 (Nippon InterMed, Japan).

It is shown (Fig. 8) that the addition of para-benzoquinone at a concentration of up to approx. 12 μg/well had an enhancing effect on the coupling of PS 12 onto polystyrene MicroWells. Concentrations of para-benzoquinone above this level, however, decreased the coupling. The binding of PS12 resisted successive treatments with 0.1% Sodium dodecyl sulphonate (SDS), 1 M acetic acid and 5 M NaCl each for 1.2 hour.

EXAMPLE 11

50 Coupling of biotin-N-hydroxy succinimide ester (NHS-d-biotin) to polystyrene solid phases modified with N,N -dimethyl hexane diamine-8-propyloxy psoralen (PS14)

A working solution of PS14



(500 μ g/ml PBS, pH 8.2, with the formula 2 M NaCl) was made from a PS14 stock solution (10 mg/ml DMSO).

100 μ l PS14 working solution was added to each well of Nunc-Immuno Module C12 (A/S NUNC, Denmark), and irradiated for 1 hour at a distance of 20 cm from an UV light source (Philips, type TL20W/09N).

Each well was washed 3 times with demineralised water, NHS-d-biotin (Hoechst, lot No. 410074, Calbiochem) was then added in a 2-fold dilution series with start concentration of 125 μ g/ml carbonate buffer, pH 9.6, 0.005 M, (1.59 g Na_2CO_3 , 2.93 g NaHCO_3 , to 1000 ml distilled water).

The MicroWell modules were incubated for two hours at room temperature. Then each well was washed 3 times with washing solution as described previously.

The coupling efficiency of biotin was visualized using avidin and horse radish peroxidase conjugated avidin mixture as described above. The substrate reaction was read using Jasco's Immuno Reader NJ-2000.

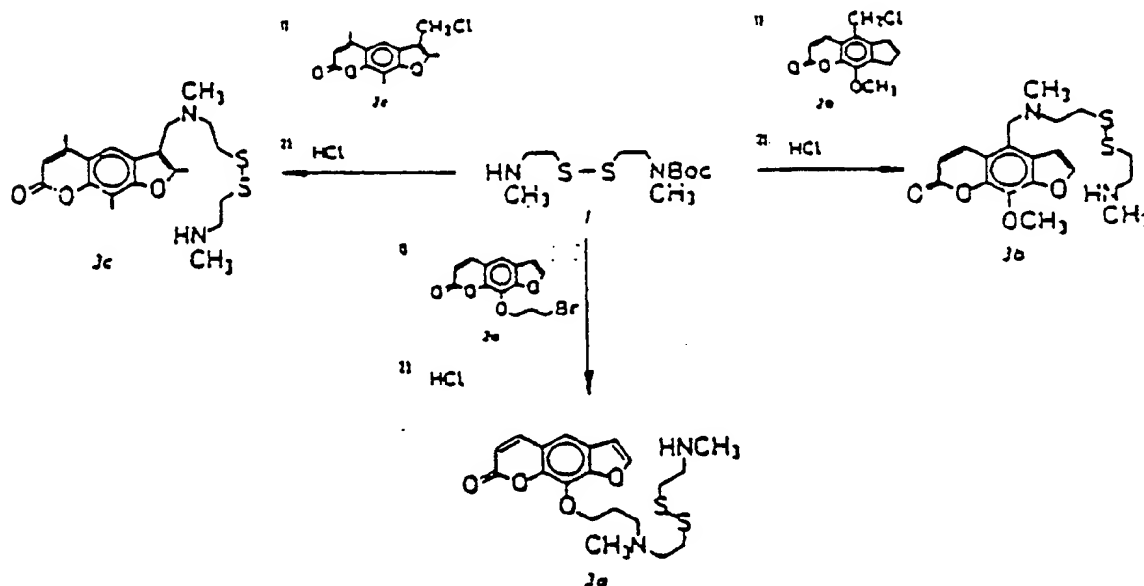
The result is shown in Fig. 9, and as can be seen, there is an excellent correlation between the amount of NHS-d-biotin added and the signal level obtained, indicating that a coupling of NHS-d-biotin to PS 14 has taken place.

This is further indicated by the fact that the addition of NHS-d-biotin with a concentration of 125 μ g/ml to non-modified wells, but otherwise treated as above, showed a mean signal level of only 0,077 OD units. The mean background signal of PS 14 was 0,094 OD units.

The binding PS14 resisted treatments with 0.1 % SDS, 1 M acetic acid and 5 M NaCl.

Preparation of formula-II compounds

The preparation of some compounds of the general formula II bound to psoralen is illustrated in the following EXAMPLES A-C. The below scheme shows the compounds prepared.



EXAMPLE A

N,N'-Dimethyl-N-[3-(8-psoralenyloxy)propyl]cysteamine (3a)

N-Tert-butoxycarbonyl-N,N'-dimethylcysteamine (1) (0.5 g, 1.6 mmol), 3-bromopropoxy-psoralen (Antonello, C., Magno, S.M., Gia O., Carlassare, F., Baccichetti, F., and Bordin, F.: *Il Farmaco*, Ed. Sci. 34, 1979, p. 139) (2a) (0.4 g, 1.6 mmol) and K_2CO_3 (0.5 g) were mixed in $CHCl_3$ (20 ml). The mixture was stirred for 72 hours and subsequently evaporated to dryness. Chromatography of the residue on silica gel with 5% methanol in methylene chloride as eluent yielded N-Boc-N,N'-dimethyl-N'-[3-(psoralenyloxy)-propyl]cysteamine (0.46 g, 60%), of which 0.22 g was dissolved in acetic acid saturated with hydrogen chloride (20 ml). After 40 minutes at room temperature compound 3a, 2 HCl, 2 H_2O precipitated.

Analysis:

Calculated for $C_{20}H_{26}N_2O_4S_2$, 2 HCl, 2 H_2O :					
	C 45.20	H 6.07	N 5.27	S 12.06	Cl 13.34
Found:	C 45.03	H 5.25	N 5.29	S 12.45	Cl 13.75.

IR-Spectrum (KBr): ν_{max} = 2900 and 3000 (NH), 1720 (CO), 1590 cm^{-1} .

Mass spectrum EI/MS, m/e: 422 (M).

The 1H NMR spectrum was in excellent agreement with the assigned structure.

EXAMPLE B

N,N'-Dimethyl-N-[(8-methoxy)psoralen-5-yl]-methyl]cysteamine(3b)

N-Boc-N,N'-dimethylcysteamine (1) (0.7 g, 2.4 mmol), 5-chloromethyl-8-methoxypsoralen (Aboulezz, A.F., El-Attar, A.A., and El-Sockary, M.: *Acta Chim. Acad. Sci. Hung.* 77, 1973, p. 208) (2b) (0.65 g, 2.4 mmol) and K_2CO_3 (0.5 g) were mixed DMF (25 ml). The mixture has heated to 70 °C for 3 hours and subsequently evaporated to dryness. Chromatography of the residue on silica gel with ethyl acetate/toluene (5-25% linear gradient) as eluent yielded N-Boc-N,N'-Dimethyl-N-[(8-methoxy)-psoralen-5-yl)methyl]-cysteamine (0.63 g, 51%). This compound (0.45 g) was deprotected as described above for compound 3a. The addition of ethyl acetate caused precipitation of 3b, 2 HCl, 1.2 H_2O (0.3 g, 70%), m.p. 208-210 °C.

Analysis:

Calculated for $C_{19}H_{24}N_2O_4S_2$, 2HCl, 1/2 H_2O :					
	C 46.53	H 5.55	N 5.71	S 13.07	Cl 14.46
Found:	C 46.35	H 5.51	N 5.42	S 12.62	Cl 14.17

IR-Spectrum (KBr): ν_{max} = 2500 and 3000 (NH), 1720 (CO), 1590 cm^{-1} .

Mass spectrum EI/MS, m/e: 408 (M).

The NMR spectrum was in excellent agreement with the assigned structure.

EXAMPLE C:

N,N'-Dimethyl-N-[(4,8,5'-trimethyl)psoralen-4'-yl)methyl]cysteamine(3c)

N-Boc-N,N'-Dimethylcysteamine (1) (0.4 g, 1.4 mmol), 4'-chloromethyl-(4,8,5'-trimethyl)psoralen (Isaacs, S.T., Shen, J.K., Hearst, J.E., and Papoport, H.: Biochemistry 16, 1977, p. 1058) (2c) (0.4 g, 1.4 mmol) and K₂CO₃ (0.5 g) were mixed in CHCl₃ (30 ml). The mixture has stirred for 27 hours and subsequently evaporated to dryness. Chromatography on silica gel with CHCl₃ as eluent yielded N-Boc-N,N'-dimethyl-N-[(4,8,5'-trimethyl)-psoralen-4'-yl)methyl]cysteamine (0.48 g, 64%), which was deprotected as described above for compound 3a. Addition of ethyl acetate caused crystallization of 3c, 2 HCl, 2 H₂O (0.40 g, 86%), m.p. 183-186° C.

Analysis:

Calculated for C₂₁H₂₈N₂O₃S₂, 2 HCl, 2 H₂O:

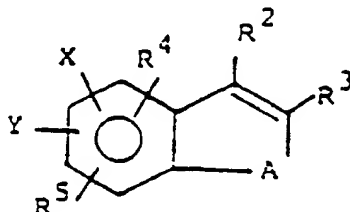
	C 47.63	H 6.47	N 5.29	S 12.11	Cl 13.39
Found:	C 47.96	H 6.57	N 5.28	S 12.05	Cl 13.33

IR-Spectrum (KBr): ν_{\max} = 2500 and 3000 (NH), 1720 (CO), 1600 cm⁻¹.
Mass spectrum FAB/MS (glycerol), m/e: 421 (M + 1).

The ¹H NMR spectrum was in excellent agreement with the assigned structure.

Claims

1. A method for modifying the surface of a polymer comprising contacting the polymer surface with a compound of the general formula I



in which A is -O-, -S-, -Se-, >NH, >NR¹, -NH-O-, -N=N-, >S⁺R¹, -S-O-, >Se + R¹, CO-O-, -CO-S-, -CS-O-, CSe-O-, -CO-Se-, -CS-NH-, -CO-NH-, -CO-N-(R¹)-, >P=O or -P(=O)(O⁻)-;

R¹ is hydrocarbyl or hydrocarbyloxy, any of which may be substituted with NO, NO₂, SO₃, CN, OH, -O, SH, SeH, PO₃⁻, PO₂⁻, COO-, halogen, epoxide, NH₂, NHR¹ or NR¹R¹ (in which R¹ has the same meaning as defined for R¹);

R², R³, R⁴, and R⁵, which may be the same or different, are H, halogen, NO₂, NO, SO₃⁻, CN, OH, O, SH, SeH, PO₃⁻, PO₂⁻, CO-O, epoxide, -NH₂, -NHR¹, -NR¹R¹ or heterocyclyl, or has the same meaning as defined for R¹;

X and Y, which may be the same or different, have the same meaning as defined for R²-R⁵, or

X and Y are adjacent to one another and together form a group with the formula -CR⁶=CR⁷-A' - where A' has the same meaning as defined for A above, and R⁶ and R⁷ which may be the same or different have the same meaning as defined for R²-R⁵; and

optionally, one or more of R¹-R⁷ designates a probe;

and irradiating the polymer and the compound of the formula I with electromagnetic radiation.

2. A method as claimed in claim 1 wherein the radiation is performed at a wavelength shorter than 700 nm, preferably shorter than 400 nm.

3. A method as claimed in claim 1 and b wherein the radiation is performed using an aqueous solution of compound I preferably having a pH of from about 5 to about 9 and optionally containing an ionic strength modifying compound preferably NaCl.

4. A method as claimed in any one of claim 1 and 3 wherein the radiation is performed in the presence of an activator molecule, preferably benzoquinone.

5. A method as claimed in claim 1 wherein the polymer is selected from polystyrene, polyethylene glycol terephthalate, polyvinyl acetate, polyvinyl chloride, polyvinyl pyrrolidone, polyacrylonitrile, polymethyl methacrylate, polytetrafluoroethylene, butyl rubber, styrene-butadiene rubber, natural rubber, polyethylene, and polypropylene.

6. A method as claimed in any of claims 1 to 5 wherein the compound I is selected from the group consisting of optionally substituted coumarins, benzofurans, indols and angelicins.

7. A method as claimed in claim 6 wherein the optionally substituted coumarins are optionally substituted psoralens.

8. A method as claimed in claim 7 wherein at least one of the substituent groups in the psoralen is an amino group containing constituent.

9. A method as claimed in claim 8 wherein the amino group containing constituent is 3-trimethylamino propoxy.

10. A method as claimed in claim 7 wherein the optionally substituted psoralen is selected from psoralen, 8-methyl-3-carboxypsoralen, 4,5',8-trimethylpsoralen, 3'-trimethyl-amino-8-propyloxy psoralen or N,N'-dimethylhexanediamine-8-propyloxypsoralen.

11. A method as claimed in claim 6 wherein the optionally substituted coumarin is warfarin.

12. A method as claimed in claim 6 wherein the optionally substituted indol is 5-hydroxytryptamine or 3-indolacetic acid.

13. A method as claimed in claim 1 wherein the probe is an organic constituent which is optionally connected to the compound I via a linking unit.

14. A method as claimed in claim 13 wherein the probe is selected from label moieties which are capable of being identified, compounds comprising reactive chemical groups, non-covalent interacting chemical groups, and other molecules.

15. A method as claimed in claim 14 wherein the label moieties which are capable of being identified are selected from the group consisting of biotin, radioactive compounds, spin labels, fluorogenic compounds, enzymes, enzyme substrates, pH colourigenic compounds, and haptens.

16. A method as claimed in claim 14 wherein the compounds comprising reactive chemical groups are selected from the group consisting of active esters capable of forming covalent bonds with other chemical groups, active halogen-containing compounds, and disulphide-containing compounds.

17. A method as claimed in claim 14 wherein the non-covalent chemical groups are charged or hydrophobic chemical groups with affinity for compounds carrying an opposite charged group or a hydrophobic group.

18. A method as claimed in claim 14 wherein the other molecules are selected from the group consisting of proteins, peptides, pharmaceuticals, mono-, oligo- or polysaccharides, or -nucleic acids, and optionally substituted lipids.

19. A method as claimed in claim 1 wherein the polymer is polystyrene, the compound of formula I is 8-propoxypsoralen which carries a probe, which is biotin connected to the compound I via N,N'-dimethyl hexane diamine linker.

20. A method as claimed in claim 1 wherein the polymer is polyvinyl chloride, the compound of formula I is 8-propoxypsoralen which carries a probe, which is biotin connected to the compound I via a N,N'-dimethyl hexane diamine linker.

21. A method as claimed in claim 1 wherein the polymer is polyethylene glycol terephthalate, the compound of formula I is 8-propoxypsoralen which carries a probe, which is biotin connected to the compound I via a N,N'-dimethyl hexane diamine linker.

22. A method as claimed in claim 1 wherein the polymer is polystyrene and the compound I is 3'-trimethyl-propyloxy-8-psoralen.

23. A modified polymer as prepared by the method of claim 1.

24. A modified polymer as claimed in claim 23 and presented in the form of a plate, a particle, a test tube or a test strip.

25. A method of determining the presence of a molecule in a sample, comprising contacting a sample containing said molecule with a modified polymer as claimed in claim 23 or 24.

26. A method as claimed in claim 25 wherein the solid surface is selected from a polymeric plate such as a thin layer plate or micro-titre-plate, polymeric beads, strips, dipsticks, test tubes, and (micro)spheres.

27. A method of immobilizing a molecule on a solid surface comprising the steps of

a) reacting the molecule with a compound of the general formula I, optionally by means of a linking unit, contacting the combination with a polymer material and subjecting the polymer and combined molecule and compound I to electromagnetic irradiation or

5 b) modifying the surface of a polymer by contacting it with a compound I as defined in claim 1 by means of electromagnetic irradiation and thereafter reacting the modified polymer surface with the molecule, optionally by means of a linking unit.

10

15

20

25

30

35

40

45

50

55

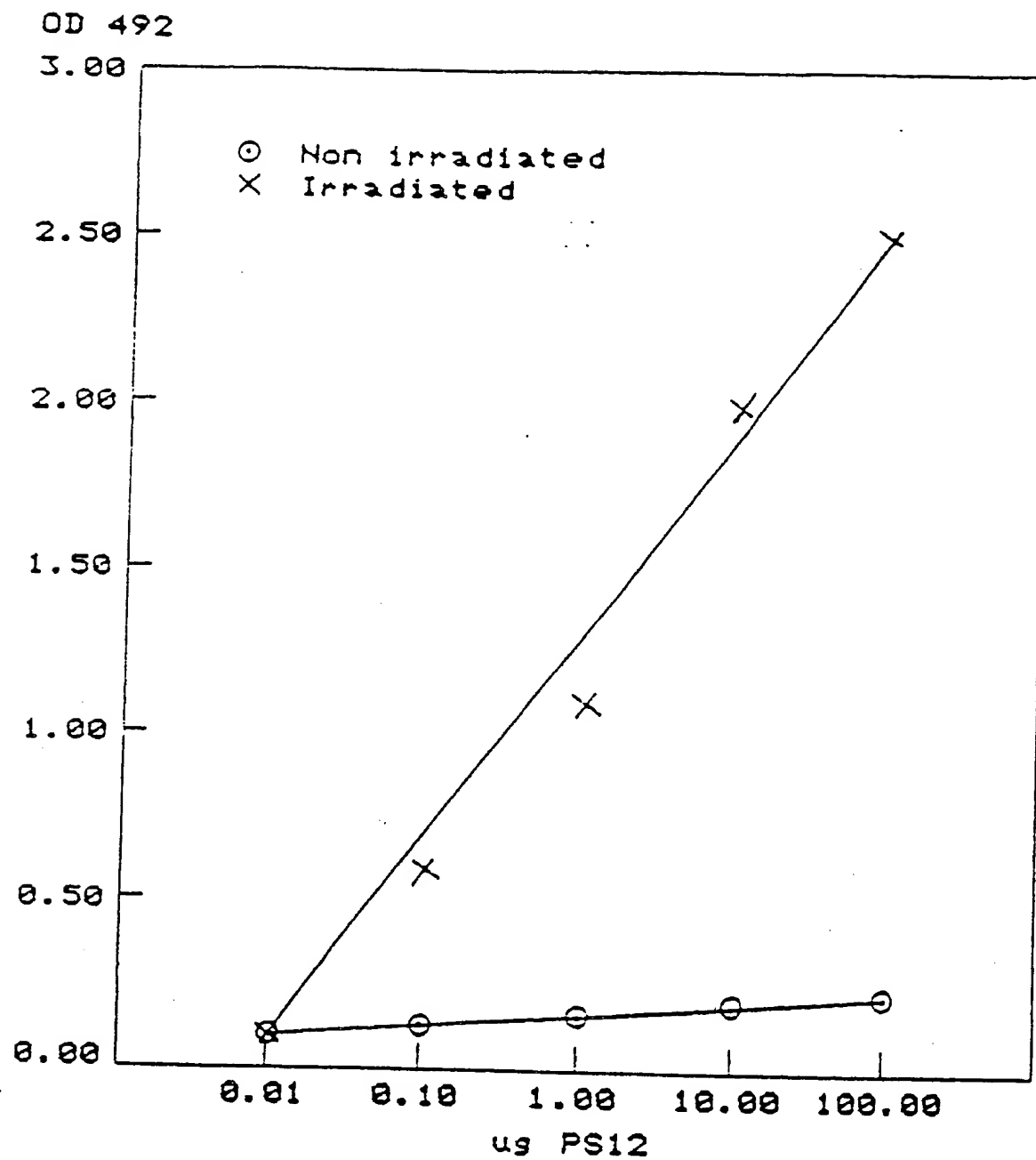


Fig. 1

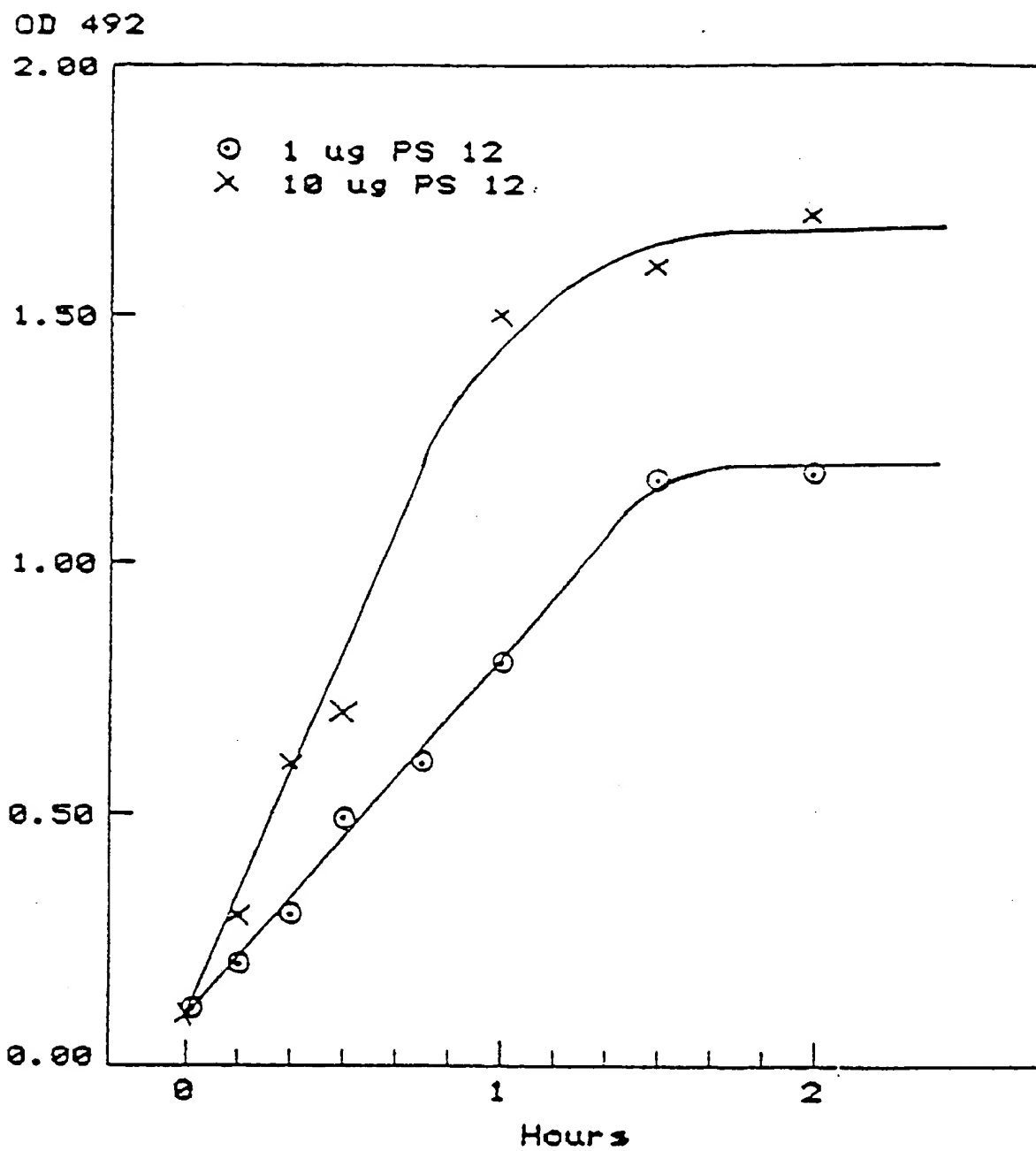


Fig. 2

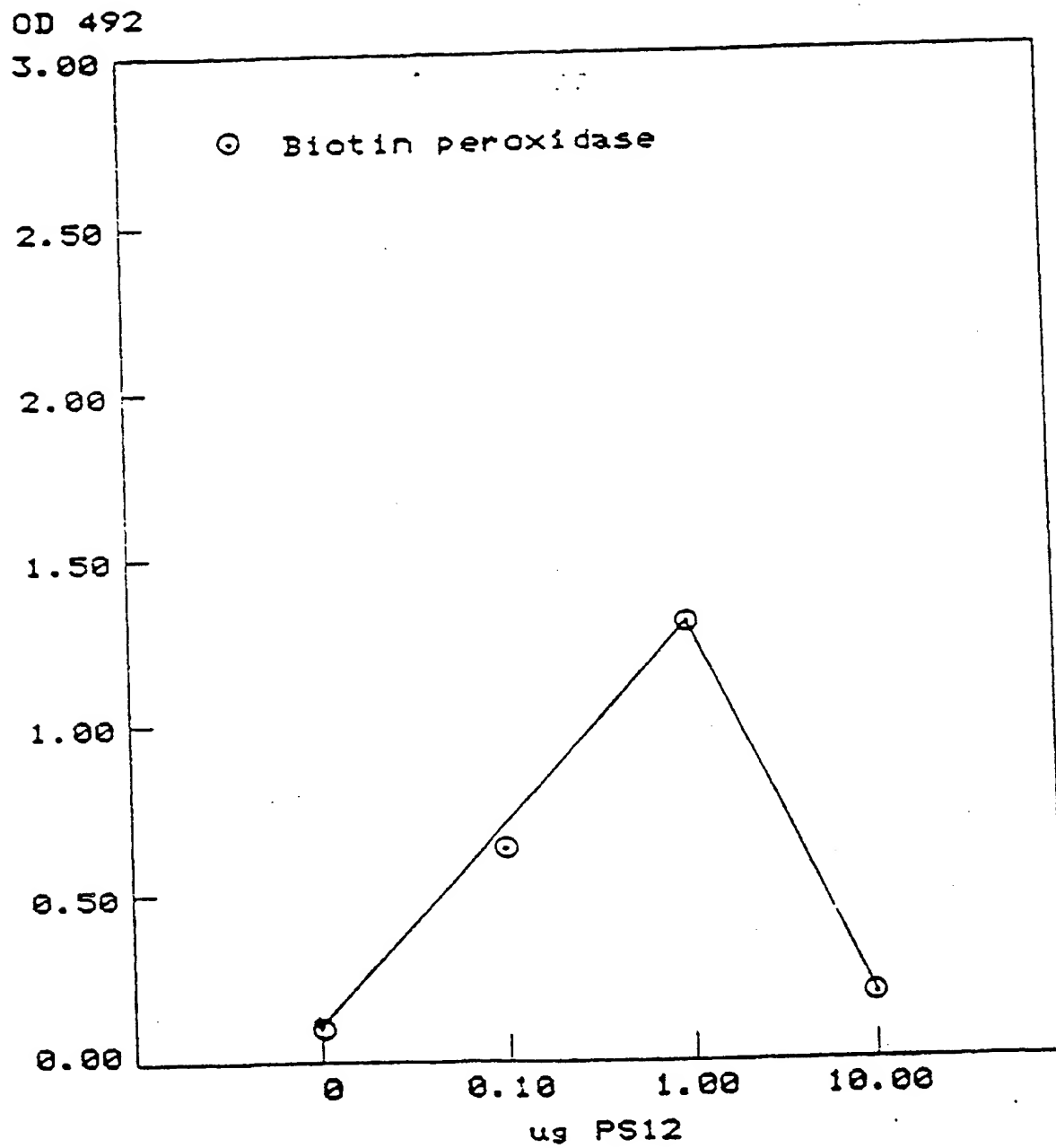
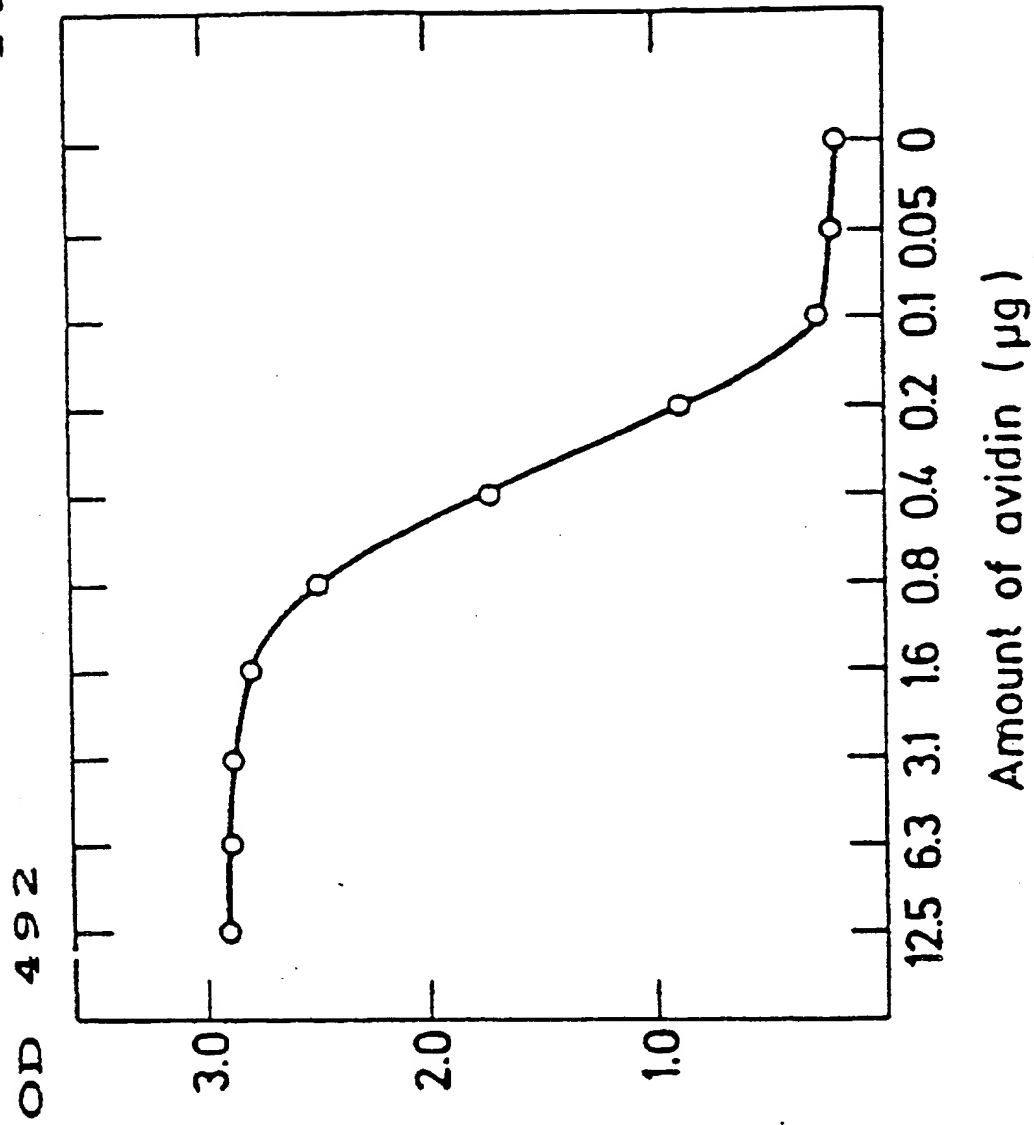
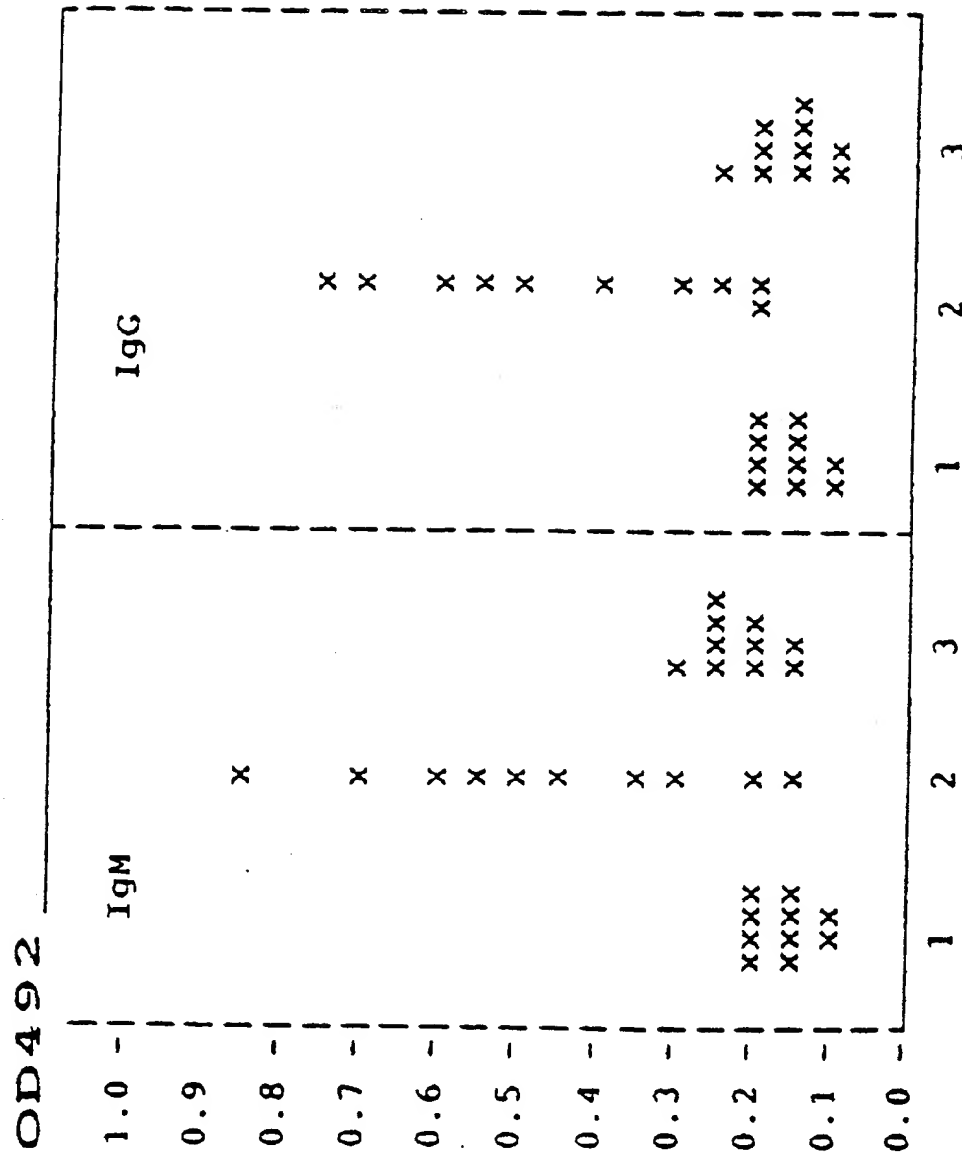


Fig. 3

Fig. 4





1: BSA
2: Avidin
3: Psoralen/biotin/streptavidin

Fig. 5

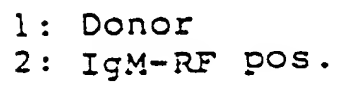
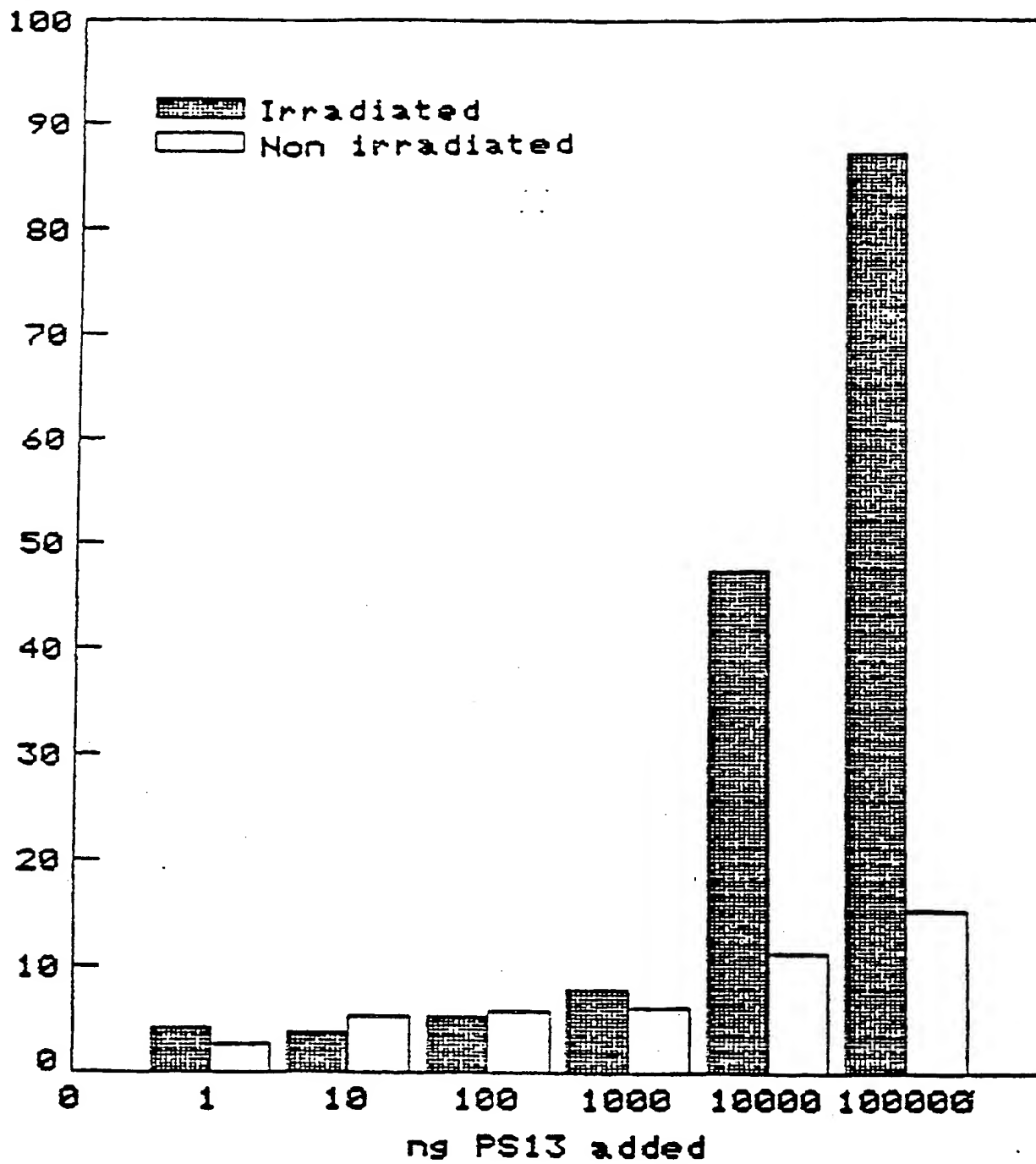
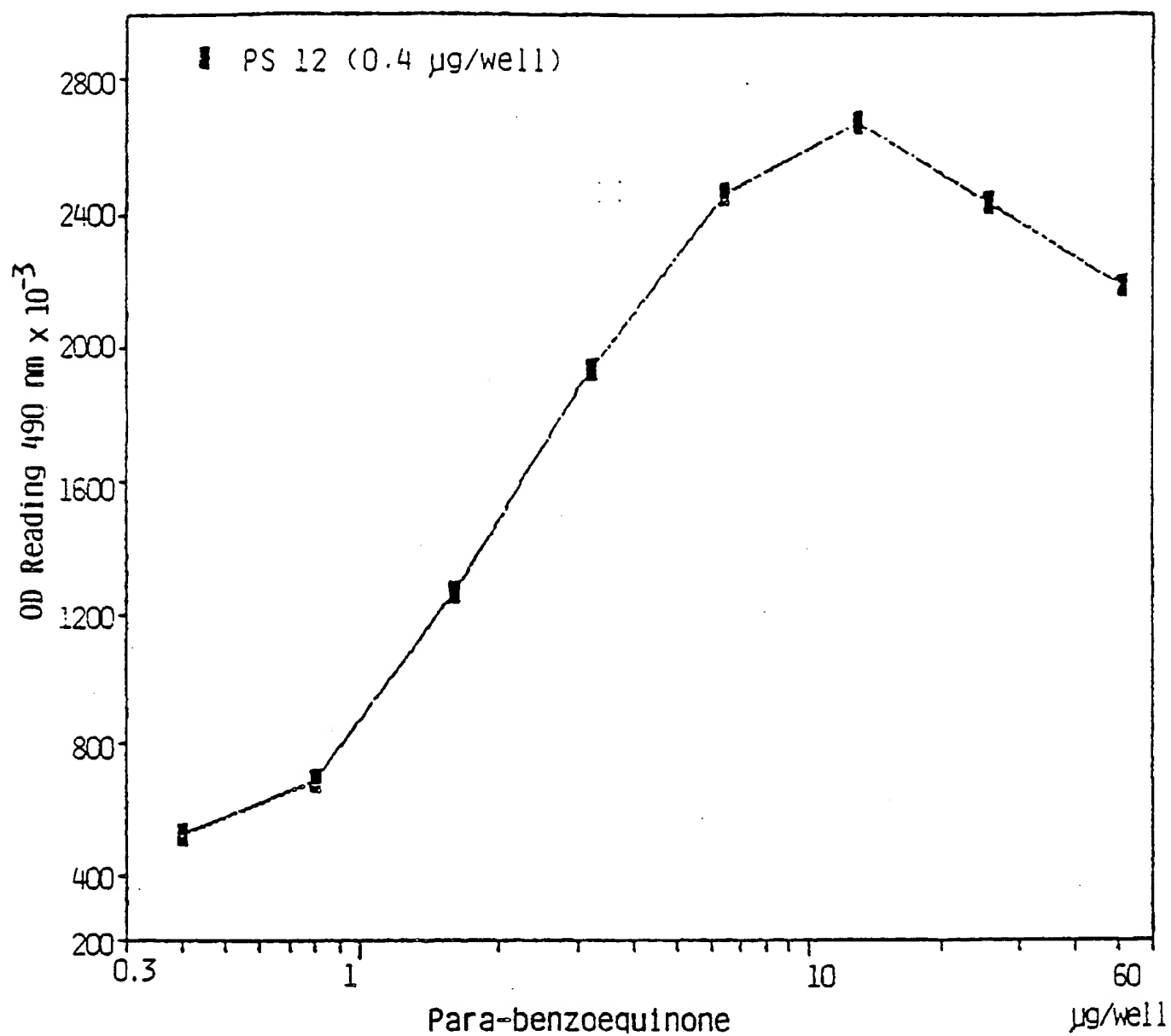


Fig. 6

EP 0 319 957 A2
Fig. 7

ng PS13 immobilized



*FIG. 8*

EP 0 319 957 A2

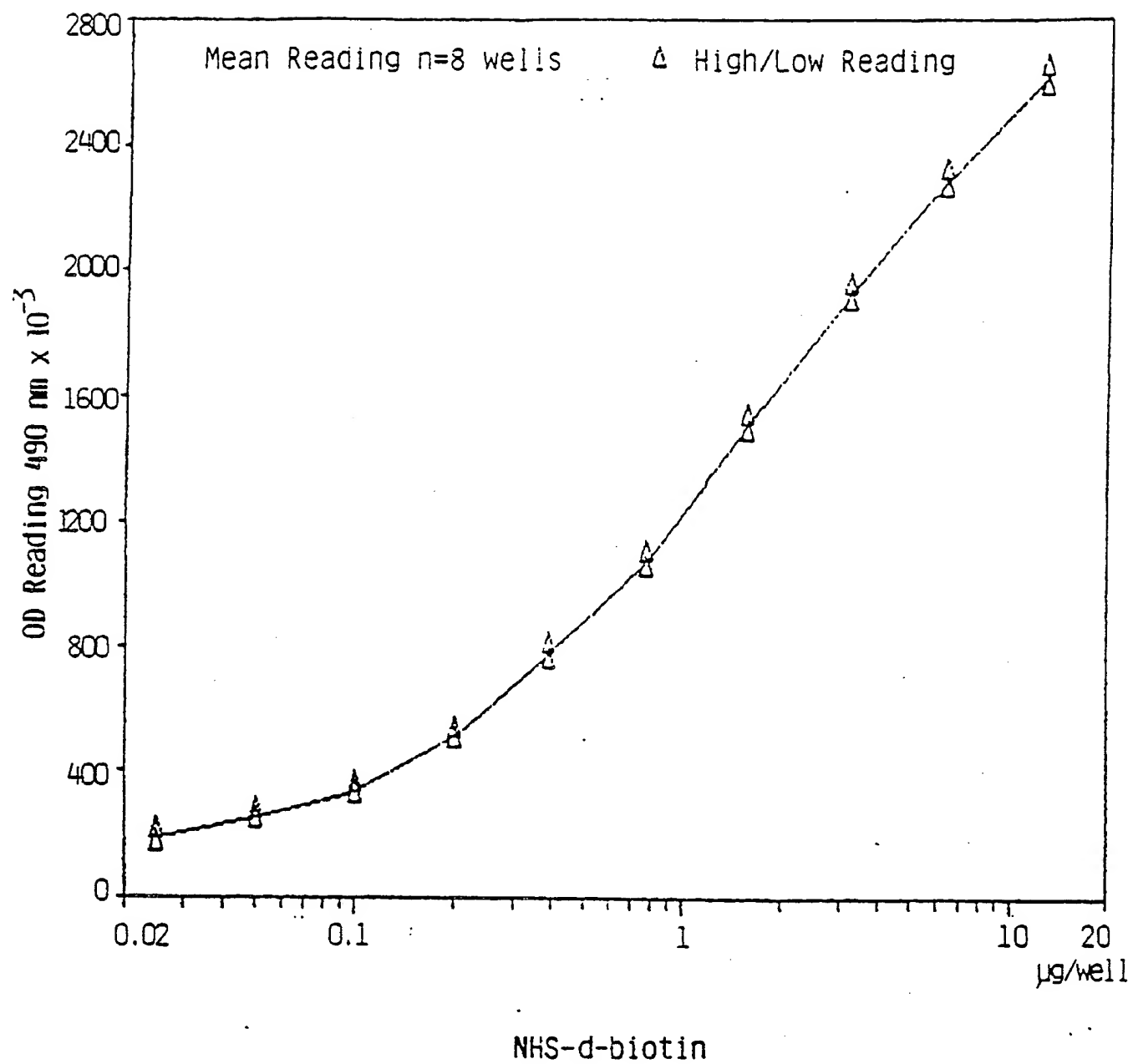


FIG. 9



Europäisches Patentamt
European Patent Office
Office européen des brevets

(11) Publication number:

**0 319 957
A3**

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 88120466.3

(51) Int. Cl.⁵: **C08J 7/12, C12N 11/00,
G01N 33/543, C12Q 1/68**

(22) Date of filing: 07.12.88

(30) Priority: 07.12.87 DK 6414/87

(43) Date of publication of application:
14.06.89 Bulletin 89/24

(84) Designated Contracting States:
ES GR

(88) Date of deferred publication of the search report:
27.12.90 Bulletin 90/52

(71) Applicant: **GLUETECH APS**
Copenhagen Science Park Symbion
Haraldsgade 68
DK-2100 Copenhagen 0(DK)

(72) Inventor: **Elsner, Henrik**
Svend Gongsvej 36
DK-2700 Bronshøj(DK)
Inventor: **Mouritsen, Søren**
Vandtaarnsvej 7, lejl. 81
DK-3460 Birkerød(DK)

(74) Representative: **Christiansen, Ejvind et al**
HOFMAN-BANG & BOUTARD A/S Adelgade
15
DK-1304 Copenhagen K(DK)

(54) **A method for modifying the surface of a polymer.**

(57) The surface of a polymer may be modified by contacting the polymer with a two or three membered heterocyclic compound I under electromagnetic irradiation. The heterocyclic compound may be substituted and may comprise linking units and/or probes so as to render the combination of polymer and compound I able to bind biomolecules and/or to be identified. Compounds I are preferably substituted psoralens, and the modified polymers can be used in e.g. ELISA methods, bioreactors, chromatography, solid phase chemical synthesis or bio-sensors.

EP 0 319 957 A3



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number

EP 88 12 0466

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
X	WO-A-8 705 805 (THE TRUSTEES OF COLUMBIA UNIVERSITY) * Page 6, lines 15-23; claim 6 * ---	1-27	C 08 J 7/12 C 12 N 11/00 G 01 N 33/543 C 12 Q 1/68
X	EP-A-0 130 523 (MOLECULAR DIAGNOSTICS INC.) * Abstract; page 3, lines 9-31; page 6, line 23 - page 7, line 5 * ---	1-27	
X	EP-A-0 281 927 (MOLECULAR DIAGNOSTICS INC.) * Page 5, lines 20-50; page 6, lines 16-20,47 - page 7, line 20 * -----	1-27	
			TECHNICAL FIELDS SEARCHED (Int. Cl.4)
			G 01 N
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 12-10-1990	Examiner CARTAGENA Y ABELLA P.
CATEGORY OF CITED DOCUMENTS			
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document		T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document	

EPO FORM 1503 03.82 (P0401)